| 1 | The Molar Ratio of Total Branched-chain Amino Acids to Tyrosine Predicts a Digit |
|----|---|
| 2 | Symbol Test Abnormality in Cirrhotic Patients |
| 3 | |
| 4 | Yusuke Mizuno ^a , Tomohisa Ishikawa ^b , Jinya Ishida ^b , Akemi Kobayashi ^c , Yasuko Konakahara ^c , |
| 5 | Akiyoshi Kinoshita ^a , Hironobu Hama ^c , Atsushi Hokari ^d , Masayuki Saruta ^b |
| 6 | |
| 7 | ^a Department of Gastroenterology and Hepatology, The Jikei University Daisan Hospital, 4-11- |
| 8 | 1 Izumihon-cho, Komae-shi, Tokyo, 201-8601, Japan |
| 9 | ^b Division of Gastroenterology and Hepatology, Department of Internal Medicine, The Jikei |
| 10 | University School of Medicine, 3-19-18 Nishi-shimbashi, Minato-city, Tokyo, 105-8471, Japan |
| 11 | ^c Clinical Nutritional Supports, Jikei University Hospital, 3-19-18 Nishi-shimbashi, Minato- |
| 12 | city, Tokyo, 105-8471, Japan |
| 13 | ^d Department of Gastroenterology and Hepatology, The Jikei University Katsushika Medical |
| 14 | Center, 6-41-2 Aoto, Katsushika-city, Tokyo, 125-8506, Japan |
| 15 | |
| 16 | Running title: BTR detects the digit symbol test abnormality. |
| 17 | |
| 18 | E-mail addresses of all co-authors |
| 19 | Tomohisa Ishikawa: ishito@jikei.ac.jp |
| | |

| 20 | Jinya | Ishida: | jikei520208 | @gmail.com |
|----|-------|---------|-------------|------------|
|----|-------|---------|-------------|------------|

- 21 Akemi Kobayashi: akobayashi@jikei.ac.jp
- 22 Yasuko Konakahara: konakahara@jikei.ac.jp
- 23 Akiyoshi Kinoshita: aki.kino@jikei.ac.jp
- 24 Hironobu Hama: hiro_hama@jikei.ac.jp
- 25 Atsushi Hokari: hokari_a@jikei.ac.jp
- 26 Masayuki Saruta: m.saruta@jikei.ac.jp
- 27
- 28 Corresponding Author: Yusuke Mizuno
- 29 Department of Gastroenterology and Hepatology, The Jikei University Daisan Hospital, 4-11-
- 30 1 Izumihon-cho, Komae-shi, Tokyo, 201-8601, Japan
- 31 Tel: +81-3-3480-1151
- 32 Fax: +81-3-3480-6688
- 33 E-mail: ymizuno0319@jikei.ac.jp
- 34

Authors' Contributions: Yusuke Mizuno and Tomohisa Ishikawa coordinated the conception and design of this study. Yusuke Mizuno, Jinya Ishida, Akemi Kobayashi and Yasuko Konakahara collected assembly, nutritional, and clinical data. Yusuke Mizuno and Tomohisa Ishikawa conducted the data analysis and drafted the manuscript. Akiyoshi Kinoshita, Hironobu Hama, Atsushi Hokari, and Masayuki Saruta performed the revision of the
manuscript. All authors read and approved this final manuscript.

Objectives: We aimed to investigate the association between the digit symbol test (DST) and 43clinical characteristics, including the nutritional status of LC patients. Methods: Fifty-nine 44cirrhotic patients without a history of overt hepatic encephalopathy were retrospectively 45evaluated. We examined neuropsychological abnormalities (NPAs) using the DST. We also 4647estimated the detailed nutritional status using the Food Frequency Questionnaire (FFQ). The patients were divided into two groups according to their DST status: patients with normal DST 48scores (DST-Nor group, n=45) and those with abnormal DST scores (DST-Abn group, n=14). 49The clinical and nutritional findings of the two groups were compared. Results: Overall, 14 50(23.7%) patients had a DST abnormality. There were significant differences between the two 51groups in serum albumin (Alb; p=0.0043), valine (Val; p=0.0016), leucine (Leu; p=0.0078), 52isoleucine (Ile; p = 0.0022), the molar ratio of total branched-chain amino acids to tyrosine 53(BTR; p=0.00025), total-bilirubin (T-Bil; p=0.0071), prothrombin time(%) (PT; p=0.028), and $\mathbf{54}$ serum sodium (Na; p=0.035). A multivariate analysis found the BTR to be the only independent 55predictor of a DST abnormality (hazard ratio, 9.24; p<0.031). An FFQ analysis, revealed that 56the nutritional findings of patients with and without a DST abnormality, were similar. 57Conclusion: The BTR was useful for predicting the risk of NPAs, as defined by a DST $\mathbf{58}$ abnormality. The risk of NPAs may be estimated by monitoring the BTR. 59

INTRODUCTION

| 62 | Various symptoms and clinical findings are indicative of the progression of liver cirrhosis (LC). |
|----|---|
| 63 | Among these symptoms, the development of hepatic encephalopathy (HE) is related to a poor |
| 64 | prognosis in patients with LC (1-3). Minimal hepatic encephalopathy (MHE), which causes |
| 65 | cognitive problems in LC patients, impairs the patient's ability to perform activities of daily |
| 66 | living and their quality of life. MHE is an early stage of overt hepatic encephalopathy (OHE) |
| 67 | (4-6). The serum ammonia level is commonly measured, and electroencephalography and |
| 68 | magnetic resonance spectroscopy are used to evaluate patients with MHE (7). |
| 69 | Some researchers have reported that the results of neuropsychiatric tests (NPTs) are |
| 70 | useful for accurately diagnosing MHE (8). The Committee of the Working Party at the Eleventh |
| 71 | World Congress of Gastroenterology Vienne (WCOG), the International Society for Hepatic |
| 72 | Encephalopathy and Nitrogen Metabolism (ISHEN), and the Japanese Society of Hepatology |
| 73 | (JSH) have recommended the use of NPTs for diagnosing MHE (8-10). The WCOG has |
| 74 | recommended that at least two of four tests (Number Connection Test-A [NCT-A], Number |
| 75 | Connection Test-B [NCT-B], Digit Symbol Test [DST] and Block Design Test [BDT]) be used |
| 76 | for the diagnosis of MHE (8, 10). In previous studies, MHE was diagnosed when at least two |
| 77 | test results were abnormal (11, 12). In other studies, MHE was diagnosed when the result of at |
| 78 | least one of the two tests was abnormal (13, 14). Among the several NPTs available, the DST |
| 79 | has been reported to have a high sensitivity and specificity for detecting neuropsychological |

| 80 | problems (14-16). However, the association between the DST and clinical characteristics, |
|----|--|
| 81 | including the nutritional status, of LC patients has not been fully clarified. |
| 82 | In the present study, we investigated the association between the DST and clinical |
| 83 | characteristics, including the nutritional status of LC patients. |
| 84 | |

85 MATERIALS AND METHODS

86 Study design

Fifty-nine cirrhotic patients without a history of OHE who had been treated at the Division of 87 Gastroenterology and Hepatology, the Jikei University School of Medicine between May 2014 88 and November 2016 were enrolled in this study. All medical records were retrospectively 89 90 reviewed for the patients' demographic and clinical data, including the nutritional status. Patients whose condition was complicated with mental disorders, renal dysfunction, bacterial 91infection, gastrointestinal hemorrhaging, or severe constipation or those treated with an 92anticonvulsant drug that might affect the cognitive function were excluded from the analysis. 93 LC was diagnosed based on physical findings, laboratory data, and clinical imaging 94characteristics. Irregularity and deformity of the shape of the liver was detected via 95ultrasonography and computed tomography. 96 The present study was approved by the institutional review board of Jikei University 97 School of Medicine (26-006 7511) and complied with the provisions of the Good Clinical 98 Practice guidelines, the Declaration of Helsinki, and local laws. All enrolled patients provided 99 100 their written informed consent for participation in the present study. 101

102 *Physical status and laboratory findings*

103 Demographic and clinical data were extracted or calculated from patient medical records. The

| 104 | body mass index (BMI) and ideal body weight were calculated. According to the assessment |
|-----|--|
| 105 | criteria for sarcopenia in liver cirrhosis established by the JSH, sarcopenia associated with liver |
| 106 | disease is known reduce the muscle mass and muscle strength. The skeletal muscle mass index |
| 107 | (SMI), calculated by dividing the left–right sum (cm ²) of the long axis (cm) \times short axis (cm) |
| 108 | of the iliopsoas muscles at the level of third lumbar vertebra by the height squared (m^2) (the |
| 109 | so-called "simple method"), correlated well with the SMI calculated using a muscle mass |
| 110 | measurement software program (cut-off value of the SMI calculated by the simple method: 6.0 |
| 111 | cm^2/m^2 in men, 3.4 cm^2/m^2 in women) (17). While we have no data on the handgrip strength |
| 112 | in our LC patients, we evaluated the SMI using a simple method for the evaluation of the |
| 113 | muscle mass. |
| 114 | Hematological and biochemical tests were performed after fasting for more than 8 h. |
| | |

The hematological analyses included the white and red blood cell counts, and platelet counts;
in addition, the hematocrit and hemoglobin levels were determined. The levels of serum alanine
transaminase (ALT), serum albumin (Alb), total bilirubin (T-Bil), creatinine, serum ammonia
(NH₃), branched-chain amino acids to tyrosine ratio (BTR), prothrombin time (PT%), and Creactive protein (CRP) were determined. As an amino acids analysis, the levels of valine (Val),
leucine (Leu), isoleucine (Ile), methionine (Met), tyrosine (Tyr), phenylalanine (Phe), histidine
(His), tryptophan (Trp), ornithine (Orn), lysine (Lys), and arginine (Arg) were also determined.

The DST was performed using a tablet-type device developed by Otsuka Pharmaceutical (Tokyo, Japan), Kokuyo (Osaka, Japan), and ISB (Tokyo, Japan); the tablet and NPT program were provided by the JSH. The DST values were estimated based on deviations from standard age- and sex-dependent values. Each patient's DST data were compared with the 10th and 90th percentile cut-off values for healthy individuals, with 5-year quartile ranges (5, 18). The DST is performed after several days of alcohol abstinence for patients with alcoholic liver cirrhosis.

131 *The evaluation of the nutritional status*

We calculated the total daily energy expenditure (TEE) using the Harris-Benedict equation. 132The activity factor was 1.3, and the injury/stress factor was 1.0, as enrolled patients had chronic 133liver disease but were ambulatory and were able to visit the hospital outpatient clinic by 134themselves (19, 20). We estimated the detailed nutritional status using the Food Frequency 135Questionnaire (FFQ), which was administered at the initiation of nutritional support and 136 performed under the supervision of a nationally registered dietician, according to our study 137schedule. The FFQ was also used to calculate the nutrient intake, estimated intake energy per 138day (EIE), and the usual daily energy ratio of nutrients (PFC ratio). The estimated intake 139amount of dietary fiber, n-3/n-6 unsaturated fatty acids and salt were calculated. The intake 140amount of each nutrient was estimated using a software program adapted to Japanese eating 141

| 142 | and living habits (Excel Eiyou-kun version 6.0; Kenpakusha, Tokyo, Japan). We compared the |
|-----|--|
| 143 | detailed nutritional data of our patients to the estimated energy requirements (EER), the |
| 144 | estimated average requirements (EAR), and adequate intake values (AI), as determined by the |
| 145 | Japanese Ministry of Health, Labor and Welfare (21, 22). |
| 146 | |
| 147 | Oral branched-chain amino acid (BCAA) supplementation |
| 148 | Patients ingested BCAA granules (LIVACT; EA Pharma Co., Ltd., Tokyo, Japan) that |
| 149 | contained 952 mg of L-isoleucine, 1904 mg of L-leucine, and 1144 mg of L-valine per sachet |
| 150 | or BCAA powder mix (Aminoleban EN; Otsuka Pharmaceutical, Tokyo, Japan) that contained |
| 151 | 2037 mg of L-leucine, 1922.5 mg of L-isoleucine, 1602 mg of L-valine, 242.5 mg of L-lysine, |
| 152 | 302 mg of arginine, 187.5 mg of histidine, and 73.5 mg of tryptophan per package. Patients |
| 153 | ingested one sachet after each meal or one to three packages daily. |
| 154 | |
| 155 | Statistical analyses |
| 156 | We used EZR (23), a graphical user interface for R, version 1.29, to perform the statistical |
| 157 | analysis (The R Foundation for Statistical Computing, Vienna, Austria). For individual |
| 158 | variables, all data were expressed as the mean and standard deviation (SD), unless stated |
| 159 | otherwise. Categorical variables were compared using the chi-squared test, while continuous |

160 variables were compared using the Mann–Whitney U test. To assess the factors predicting a

| 161 | DST abnormality, logistic regression analyses were used to calculate odds ratios (ORs) and |
|-----|--|
| 162 | 95% confidence intervals (CIs). Only variables with a p-values of <0.05 in a univariate analysis |
| 163 | were included in the multivariate analysis. |
| 164 | A receiver operating characteristics (ROC) was also generated to determine the optimal |
| 165 | cut-off value of continuous variables. P-value of < 0.05 were considered to indicate statistical |
| 166 | significance. |
| 167 | |

168 **RESULTS**

169 *Etiology and clinical findings*

The patient demographics and clinical characteristics are shown in Table 1. The mean age of 170the patients was 66 years old. Thirty-six (61%) patients were men, and 23 (39%) were women. 171The mean BMI was 24.0 ± 4.7 kg/m². The most common etiology of LC was hepatitis C (n=24, 17217340.7%), followed by alcohol-associated hepatitis (n=17, 28.8%), NASH (n=6, 10.1%), and hepatitis B (n=4, 6.8%). Of the 59 patients enrolled, none showed portosystemic shunt on 174dynamic-enhanced computed tomography scans or ultrasonography. Thirty-one patients 175(52.5%) had a history of hepatocellular carcinoma (HCC) treatment. Twenty-seven patients 176(45.8%) used BCAA supplementation. Thirteen patients took 1 BCAAs granule sachet after 177each meal, whereas BCAA powder mix was used by 14 patients (11 patients took 1 package 178daily as a late-evening snack, 1 patient took 2 packages per day, and 2 patients took 3 packages 179per day). The Child–Pugh stages of the patients were as follows: stage A (n=37), B (n=14), and 180 C(n=8). The mean Child–Pugh score (CP-S) was 6.8 ± 2.1 . The mean Alb (3.4 ± 0.5 g/dL), 181 BTR (3.9 ± 1.6), and platelet counts (11.5 ± $5.9 \times 10^4/\mu$ L) were lower than the reference values. 182183

184 DST results and their correlation with clinical characteristics

185 We divided patients into two groups based on the reference values (DST-Abn group, n=14,

186 DST-Nor group, n=45). The correlation between the DST status and the clinical characteristics

| 187 | is shown in Table 2 and Figure 1. The CP-S of the DST-Abn group was significantly higher |
|-----|--|
| 188 | than that of the DST-Nor group (6.3 ± 1.3 vs. 8.8 ± 2.9 , p = 0.0052). The DST-Abn group also |
| 189 | had markedly higher T-Bil levels than the DST-Nor group (2.6 ± 1.7 vs. 1.2 ± 0.5 , $p = 0.0071$). |
| 190 | In addition, the levels of Alb (3.0 \pm 0.5 vs. 3.5 \pm 0.5, p = 0.0043), PT (62.4 \pm 20.9 vs. 76.7 \pm |
| 191 | 14.3, p = 0.028), serum sodium (137.6 \pm 3.5 vs. 140.0 \pm 2.6, p = 0.035), Val (168.6 \pm 60.9 vs. |
| 192 | 226.9 ± 49.0, p = 0.0016), Leu (87.1 ± 36.2 vs. 122.3 ± 31.2, p = 0.0078), and Ile (49.5 ± 19.1 |
| 193 | vs. 69.0 \pm 17.0, p = 0.0022) as well as the BTR values (2.8 \pm 1.1 vs. 4.3 \pm 1.6, p = 0.00025) |
| 194 | were notably lower in the DST-Abn group than in the DST-Nor group. There were no |
| 195 | significant differences between the two groups regarding the use or non-use of BCAA |
| 196 | supplementation ($p = 0.776$). |

197The results of the multivariate analysis are shown in Table 3. While univariate analyses identified significant differences in CP-S, the CP-S is composed of Alb, T-Bil, PT, ascites, and 198hepatic encephalopathy and was therefore excluded from the multivariate analysis. As a result, 199a BTR of <2.92 was found to be an independent predictor of a DST abnormality (hazard ratio 200[HR], 9.24; p < 0.031). 201

Given the possible effect of BCAA supplementation on the DST and BTR, LC patients 202not taking BCAA supplementation were divided into two groups according to the DST result. 203The clinical characteristics of the LC patients without BCAA supplementation who had normal 204and abnormal DST results are shown in Table 6. There was a significant difference in the BTR 205

between the two groups in a univariate analysis (p = 0.0385). A multivariate analysis was not performed due to the small number of subjects.

208

209 The estimation of the nutritional status using the FFQ

Overall, 41 enrolled LC cases completed the FFQ. The remaining 18 patients were unable to 210211complete the FFQ because of the amount of time required. We estimated the nutritional status of 41 patients using physical measurements and the FFQ (Table 4). The TEE was not 212significantly different from the EIE (men: 1769.4 ± 309.3 kcal vs. 1932.7 ± 533.4 kcal, p > 2130.05; women: 1515.7 ± 149.7 kcal vs. 1636.9 ± 455.8 kcal, p > 0.05). The protein ingestion 214values (men: 1.2 ± 0.3 g/kg; women: 1.1 ± 0.4 g/kg) were not significantly different from the 215216recommended values. The fat energy ratio (men: $28.6\% \pm 5.2\%$; women: $30.2\% \pm 4.5\%$) was 217significantly higher than the AI (20.0%-25.0%). We found that 36.6% of the patients had a fat energy ratio of >30% (n=15/41). The dietary fiber intake of men (16.2 ± 6.0 g/day; vs. women, 218 16.2 ± 4.4 g/day) was significantly less than the AI (>19 g/day, p = 0.03; vs. women, >17 g/day, 219 p = 0.60). 220

221

222 The nutritional condition according to the DST status

In the 41 LC patients who completed the FFQ, we compared the nutritional condition between

the DST-Abn and DST-Nor groups. There were only 7 patients (2 men and 5 women) in the

| 225 | DST-Abn group. We therefore judged that it was impossible to consider male and female |
|-----|---|
| 226 | patients separately. Nutritionally, the DST-Abn and DST-Nor groups were similar (Table 5). |
| 227 | Based on the FFQ analysis, both groups had fat energy ratios that were above the AI, and both |
| 228 | groups demonstrated insufficient dietary fiber intake. |

230 **DISCUSSION**

HE serves as a prognostic factor in patients with LC. Although studies have reported that half of MHE patients develop OHE within three years (1, 3), the diagnostic methods and detailed pathogenesis of MHE remain to be clarified. Based on these limitations, as well as the adverse effects on the patient's quality of life, we consider the establishment of diagnostic criteria for MHE to be imperative.

Most LC patients have protein energy malnutrition (PEM) and accelerated resting energy expenditure. Drastic changes in the metabolic nutrient ratio are observed in patients with liver dysfunction, in addition to a decreased carbohydrate ratio and increased fat ratio (24, 25). The absorption of aromatic amino acids into the nerve cells is also stimulated due to an amino acid imbalance, which leads to higher-brain-function restrictions and the development of HE (26-28). Practical nutritional support, which can help overcome nutritional problems, improves the prognosis of LC patients (25, 29, 30)[.]

In the present study, we selected the DST as an indicator of earlier neurophysiological problems in cirrhotic patients. The approach used in this study conformed to the WCOG report, which requires that a neurophysiological assessment be used to diagnose MHE after excluding any other brain disorders (8). The NPTs, which are recommended by the WCOG, ISHEN, and JSH, are a valuable and useful method for diagnosing MHE (8-10). Michitaka et al. reported that abnormalities in 2 NPT items has 80% specificity for MHE, while abnormalities in \geq 3 items has 95% specificity (12). Some studies have reported that more than two abnormal test
items in the NCT-A/B, DST, and BDT are indicative of MHE (31). Li et al. reported that MHE
could be diagnosed with 76.9% sensitivity and 96.3% specificity using the combination of the
NCT-A and DST (15). Weissenberg et al. reported that, among the NPTs, the DST has high
sensitivity (80.0%) and specificity (96.5%) for the diagnosis of MHE (16). We therefore
selected the DST to evaluate early neurophysiological problems.

In our study, 14 (23.7%) patients were classified into the DST-Abn group, indicating 255that patients in this group are at risk of developing MHE. Although the CP-S in the DST-Abn 256group was higher than that in the DST-Nor group, four patients in the DST-Abn group had a 257Child-Pugh classification of A. Researchers are particularly interested in such groups of 258patients, as these patients have a relatively good clinical liver function. The values of Val, Leu, 259Ile, and the BTR in the DST-Abn group were significantly lower than in the DST-Nor group. 260Furthermore, a multivariate analysis demonstrated that a BTR <2.92 was independently 261associated with DST abnormality. Recently, Hanai et al. reported that the presence of 262sarcopenia and a low BCAAs level (<327 nmol/ml) was independently associated with MHE 263in LC patients, supporting our findings (32). Previous studies reported that the serum BCAA 264values might be remarkably decreased in patients with compensated LC in Child-Pugh class A; 265in particular, lower values were closely associated with the liver prognosis and mortality 266(33-35). The ammonia detoxification pathway includes the urea cycle in the liver and the 267

glutamine synthesis pathway in the skeletal muscle and brain. In cirrhotic patients, while the 268ability to process ammonia by the urea cycle is reduced, the detoxification of ammonia to 269glutamine in skeletal muscle, brain, and likely the lungs is activated. When glutamate and 270ammonia react to form glutamine, ammonia is processed. However, an increase in the 271concentration of glutamine induces hepatic encephalopathy by causing astrocyte swelling and 272273cerebral hypertension in the brain. Furthermore, in skeletal muscle, BCAAs (Val, Leu, Ile) are metabolized to acetyl-CoA and succinyl-CoA and converted to a-ketoglutarate in the Krebs 274cycle. α-ketoglutarate itself is also a source of glutamate, generating ammonia through the 275process of producing glutamine. However, the produced glutamine is metabolized in the 276intestine and kidney to ammonia and glutamate, and glutamate in also decomposed into a-277ketoglutarate and ammonia. Even if the ammonia concentration decreases in a certain organ, 278ammonia can be consequently produced in another organ, resulting in a vicious cycle in which 279the ammonia level in the body does not decrease (34, 36). We hypothesize that the 280neuropsychological dysfunction and MHE associated with LC is related to the fractional 281imbalance of amino acids in these patients. In fact, Akahoshi et al. demonstrated that NPT 282283scores (including those of the NCT, BDT, and DST) were negatively associated with the BTR in patients undergoing liver transplantation and that the BTR in patients with MHE was 284significantly lower than that in those without MHE (37). We therefore suggest that the BTR is 285a useful parameter for predicting the development of cognitive problems similar to MHE. 286

| 287 | The FFQ have been used in nutritional epidemiological studies to investigate the |
|-----|--|
| 288 | association between food and chronic diseases (38-44). Evaluating the dietary intake helps |
| 289 | physicians and dieticians understand poor food habits, dietary choice, and excessive |
| 290 | consumption. Suitable management of malnutrition involves establishing daily a diet that |
| 291 | includes a restricted calorie intake and diverse nutrient consumption (45). Using the FFQ, the |
| 292 | quantity of daily intakes was calculated based on the portion size and consumption frequency. |
| 293 | The TEE/EIE ratios in men and women in the present study were 1.0 \pm 0.5 and 1.0 \pm 0.3, |
| 294 | respectively. The EIE as estimated by the FFQ and the TEE as calculated by the Harris- |
| 295 | Benedict equation were almost same value. Furthermore, the EIE/BW and protein intake in this |
| 296 | study were similar to the ideal values established in the European Society for Parenteral and |
| 297 | Enteral Nutrition (ESPEN) guidelines and in the ISHEN recommendations (29, 30). |
| 298 | The ideal AI for the fat energy ratio is reported to be 20%-25% (21, 22). In the present |
| 299 | study, regardless of the presence of an abnormal DST, the fat energy ratio (men: 28.6% \pm 5.2%; |
| 300 | women: $30.2\% \pm 4.5\%$) exceeded that of the ideal AI (21, 22). Consistent with our results, |
| 301 | Tajika et al. showed that the rate of fat oxidation in cirrhotic patients (47.8%), as determined |
| 302 | by an indirect calorimeter, was higher than that in the control group (26.6%) (24). In addition, |
| 303 | the lipid uptake rate of NASH patients estimated using a 4-day diet diary was reported to be |
| 304 | 35% of the total energy calories (38). The findings of these reports are similar to our results, |
| 305 | indicating the accuracy of the FFQ results in our study. |

20

| 306 | In the present study, the intake of dietary fiber by the enrolled patients was lower than |
|-----|--|
| 307 | the AI. Dietary fiber reduces gut digestion, passage time, and ammonia absorption. A previous |
| 308 | study reported a close relationship between gut flora and the ingestion of dietary fiber, with a |
| 309 | reduced fiber consumption resulting in gut flora changes (46-48). Nutrient ratios and dietary |
| 310 | fiber ingestion should be further investigated as a means of preventing cognitive problems, |
| 311 | including DST abnormalities and even HE. |
| 312 | There were no marked nutritional differences between the DST-Abn and DST-Nor |
| 313 | groups in our study. In contrast, a recent report, by Zhu et al. showed that the total protein |
| 314 | intake, total unsaturated fatty acid intake, and total carbohydrate intake were associated with |
| 315 | changes in the mean total latency and number of errors in a symbol digit substitution test (49). |
| 316 | The small sample size of our study may have affected the statistical analyses. |
| 317 | This study was associated with several limitations. First, the study was retrospective in |
| 318 | nature and included a relatively small number of patients who were managed in a single center. |
| 319 | Although the multivariate analysis showed the BTR to be an independent marker for detecting |
| 320 | a DST abnormality, this method is of limited value in a retrospective, small-scale study. These |
| 321 | methodological drawbacks may reduce the strength of our statistical conclusions. Second, we |
| 322 | did not directly compare the FFQ and the measurement results, including the oxidation rates of |

carbohydrate, fat, and protein, respiratory quotient and resting energy expenditure, as 323determined by an indirect calorimeter. Third, liver cancer may affect the nutritional status. 324

| 325 | Nevertheless, LC patients with liver cancer and a good performance status are often |
|-----|---|
| 326 | encountered in actual clinical practice and participated in the current study. Thus, a large-scale, |
| 327 | prospective study is required to confirm our findings. |
| 328 | In conclusion, we found that the pathophysiology of a DST abnormality is closely |
| 329 | related to decreases in the BTR level. The BTR is the most useful predictor of a DST |
| 330 | abnormality in LC patients. The NPA risk may be estimated by monitoring the BTR. |
| 331 | |
| 332 | ACKNOWLEDGMENTS |
| 333 | We acknowledge the members of the University Hospital Nutrition Support Team for their |
| 334 | advice and assistance. |
| 335 | |
| 336 | Conflicts of Interest and Funding Disclosure |
| 337 | The authors declare no conflict of interest in association with the present study. This research |
| 338 | did not receive any specific grant from funding agencies in the public, commercial, or not-for- |
| 339 | profit sectors. |
| 340 | |

341 **REFERENCES**

| 342 | 1. Stewart CA, Malinchoc M, Kim WR, Kamath PS. Hepatic encephalopathy as a predictor of |
|-----|---|
| 343 | survival in patients with end-stage liver disease. Liver Transpl. 2007 Oct; 13(10):1366-71. |
| 344 | 2. Bustamante J, Rimola A, Ventura PJ, Navasa M, Cirera I, Reggiardo V, et al. Prognostic |
| 345 | significance of hepatic encephalopathy in patients with cirrhosis. J.Hepatol. 1999 May; |
| 346 | 30(5):890-5. |
| 347 | 3. Garg H, Kumar A, Garg V, Sharma P, Sharma BC, Sarin SK. Clinical profile and predictors |
| 348 | of mortality in patients of acute-on-chronic liver failure. Dig.Liver Dis. 2012 Feb; 44(2):166- |
| 349 | 71. |
| 350 | 4. Romero-Gomez M, Boza F, Garcia-Valdecasas MS, Garcia E, Aguilar-Reina J. Subclinical |

- hepatic encephalopathy predicts the development of overt hepatic encephalopathy.
 Am.J.Gastroenterol. 2001 Sep; 96(9):2718-23.
- 5. Kato A, Tanaka H, Kawaguchi T, Kanazawa H, Iwasa M, Sakaida I, et al. Nutritional
 management contributes to improvement in minimal hepatic encephalopathy and quality of life
 in patients with liver cirrhosis: A preliminary, prospective, open-label study. Hepatol.Res. 2013
 May; 43(5):452-8.

6. Bajaj JS, Pinkerton SD, Sanyal AJ, Heuman DM. Diagnosis and treatment of minimal
hepatic encephalopathy to prevent motor vehicle accidents: A cost-effectiveness analysis.
Hepatology. 2012 Apr; 55(4):1164-71.

| 360 | 7. Dhiman RK, Saraswat VA, Sharma BK, Sarin SK, Chawla YK, Butterworth R, et al. |
|-----|--|
| 361 | Minimal hepatic encephalopathy: Consensus statement of a working party of the indian |
| 362 | national association for study of the liver. J.Gastroenterol.Hepatol. 2010 Jun; 25(6):1029-41. |
| 363 | 8. Ferenci P, Lockwood A, Mullen K, Tarter R, Weissenborn K, Blei AT. Hepatic |
| 364 | encephalopathydefinition, nomenclature, diagnosis, and quantification: Final report of the |
| 365 | working party at the 11th world congresses of gastroenterology, vienna, 1998. Hepatology. |
| 366 | 2002 Mar; 35(3):716-21. |
| | |

_ _ _ ~

att a1

1

- 9. Randolph C, Hilsabeck R, Kato A, Kharbanda P, Li YY, Mapelli D, et al.
 Neuropsychological assessment of hepatic encephalopathy: ISHEN practice guidelines. Liver
 Int. 2009 May; 29(5):629-35.
- 10. Kato A, Kato M, Ishii H, Ichimiya Y, Suzuki K, Kawasaki H, et al. Development of
- 371 quantitative neuropsychological tests for diagnosis of subclinical hepatic encephalopathy in
- 372 liver cirrhosis patients and establishment of diagnostic criteria-multicenter collaborative study
- 373 in japanese. Hepatol.Res. 2004 Oct; 30(2):71-8.

- ----

D 17

....

~1

- 11. Prasad S, Dhiman RK, Duseja A, Chawla YK, Sharma A, Agarwal R. Lactulose improves
- 375 cognitive functions and health-related quality of life in patients with cirrhosis who have
- 376 minimal hepatic encephalopathy. Hepatology. 2007 Mar; 45(3):549-59.

| 377 | 12. Michitaka K, Tokumoto Y, Uesugi K, Kisaka Y, Hirooka M, Konishi I, et al. |
|-----|---|
| 378 | Neuropsychiatric dysfunction in patients with chronic hepatitis and liver cirrhosis. Hepatol.Res. |
| 379 | 2008; 38(11):1069-75. |

- 13. Bajaj JS, Saeian K, Verber MD, Hischke D, Hoffmann RG, Franco J, et al. Inhibitory
- 381 control test is a simple method to diagnose minimal hepatic encephalopathy and predict
- development of overt hepatic encephalopathy. Am.J.Gastroenterol. 2007 Apr; 102(4):754-60.
- 383 14. Maric D, Klasnja B, Filipovic D, Brkic S, Ruzic M, Bugarski V. Minimal hepatic
- encephalopathy in patients with decompensated liver cirrhosis. Acta Clin.Croat. 2011 Sep;
 50(3):375-80.
- 15. Li SW, Wang K, Yu YQ, Wang HB, Li YH, Xu JM. Psychometric hepatic encephalopathy
- score for diagnosis of minimal hepatic encephalopathy in china. World J.Gastroenterol. 2013
 Dec 14; 19(46):8745-51.
- 389 16. Weissenborn K, Ennen JC, Schomerus H, Ruckert N, Hecker H. Neuropsychological
 390 characterization of hepatic encephalopathy. J.Hepatol. 2001 May; 34(5):768-73.
- 17. Nishikawa H, Shiraki M, Hiramatsu A, Moriya K, Hino K, Nishiguchi S. Japan society of
- 392 hepatology guidelines for sarcopenia in liver disease (1st edition): Recommendation from the
- working group for creation of sarcopenia assessment criteria. Hepatol.Res. 2016 Sep;
 46(10):951-63.

| 395 | 18. Kawaguchi T, Konishi M, Kato A, Kato M, Kooka Y, Sawara K, et al. Updating the |
|-----|---|
| 396 | neuropsychological test system in japan for the elderly and in a modern touch screen tablet |
| 397 | society by resetting the cut-off values. Hepatol.Res. 2017 Nov; 47(12):1335-9. |
| 398 | 19. Long CL, Schaffel N, Geiger JW, Schiller WR, Blakemore WS. Metabolic response to |
| 399 | injury and illness: Estimation of energy and protein needs from indirect calorimetry and |
| 400 | nitrogen balance. JPEN J.Parenter.Enteral Nutr. 1979 Nov-Dec; 3(6):452-6. |
| 401 | 20. Barak N, Wall-Alonso E, Sitrin MD. Evaluation of stress factors and body weight |
| 402 | adjustments currently used to estimate energy expenditure in hospitalized patients. JPEN |

- 403 J.Parenter.Enteral Nutr. 2002 Jul-Aug; 26(4):231-8.
- 404 21. Tsuboyama-Kasaoka N, Takizawa A, Tsubota-Utsugi M, Nakade M, Imai E, Kondo A, et
- 405 al. Dietary intake of nutrients with adequate intake values in the dietary reference intakes for
- 406 japanese. J.Nutr.Sci.Vitaminol.(Tokyo). 2013 ; 59(6):584-95.
- 407 22. Megumi Tsubota-Utsugi, Eri Imai, Makiko Nakade, Nobuyo Tsuboyama-Kasaoka, Akemi
- 408 Morita and Shinkan Tokudome, Ph.D, M.D. Dietary reference intakes for japanese -2010-.
- 409 23. Kanda Y. Investigation of the freely available easy-to-use software 'EZR' for medical
- 410 statistics. Bone Marrow Transplant. 2013 Mar; 48(3):452-8.
- 411 24. Tajika M, Kato M, Mohri H, Miwa Y, Kato T, Ohnishi H, et al. Prognostic value of energy
- 412 metabolism in patients with viral liver cirrhosis. Nutrition. 2002 Mar; 18(3):229-34.

- 413 25. Tsiaousi ET, Hatzitolios AI, Trygonis SK, Savopoulos CG. Malnutrition in end stage liver
 414 disease: Recommendations and nutritional support. J.Gastroenterol.Hepatol. 2008 Apr;
 415 23(4):527-33.
- 416 26. Albrecht J, Norenberg MD. Glutamine: A trojan horse in ammonia neurotoxicity.
- 417 Hepatology. 2006 Oct; 44(4):788-94.
- 418 27. Plauth M, Merli M, Kondrup J, Weimann A, Ferenci P, Muller MJ, et al. ESPEN guidelines
- 419 for nutrition in liver disease and transplantation. Clin.Nutr. 1997 Apr; 16(2):43-55.
- 420 28. Muller MJ, Bottcher J, Selberg O, Weselmann S, Boker KH, Schwarze M, et al.
- 421 Hypermetabolism in clinically stable patients with liver cirrhosis. Am.J.Clin.Nutr. 1999 Jun;
 422 69(6):1194-201.
- 423 29. Plauth M, Cabre E, Riggio O, Assis-Camilo M, Pirlich M, Kondrup J, et al. ESPEN
- 424 guidelines on enteral nutrition: Liver disease. Clin.Nutr. 2006 Apr; 25(2):285-94.
- 425 30. Amodio P, Bemeur C, Butterworth R, Cordoba J, Kato A, Montagnese S, et al. The
- 426 nutritional management of hepatic encephalopathy in patients with cirrhosis: International
- 427 society for hepatic encephalopathy and nitrogen metabolism consensus. Hepatology. 2013 Jul;
 428 58(1):325-36.
- 429 31. Hirano H, Saito M, Yano Y, Momose K, Yoshida M, Tanaka A, et al. Chronic liver disease
 430 questionnaire would be a primary screening tool of neuropsychiatric test detecting minimal
- 431 hepatic encephalopathy of cirrhotic patients. Hepatol.Res. 2015 Jul 31.

| 432 | 32. Hanai T, Shiraki M, Watanabe S, Kochi T, Imai K, Suetsugu A, et al. Sarcopenia predicts |
|-----|---|
| 433 | minimal hepatic encephalopathy in patients with liver cirrhosis. Hepatol.Res. 2017 Dec; |
| 434 | 47(13):1359-67. |

- 435 33. Kinny-Koster B, Bartels M, Becker S, Scholz M, Thiery J, Ceglarek U, et al. Plasma amino
- 436 acid concentrations predict mortality in patients with end-stage liver disease. PLoS One. 2016
- 437 Jul 13; 11(7):e0159205.
- 438 34. Holecek M. Ammonia and amino acid profiles in liver cirrhosis: Effects of variables leading
- to hepatic encephalopathy. Nutrition. 2015 Jan; 31(1):14-20.
- 440 35. Kato A, Suzuki K. How to select BCAA preparations. Hepatol.Res. 2004 Dec; 30S:30-5.
- 441 36. Manoli I, Venditti CP. Disorders of branched chain amino acid metabolism. Transl.Sci.Rare
- 442 Dis. 2016 Nov 7; 1(2):91-110.
- 443 37. Akahoshi M, Ichikawa T, Taura N, Miyaaki H, Yamaguchi T, Yoshimura E, et al. Sleep
- 444 disturbances and quality of life in patients after living donor liver transplantation.
- 445 Transplant.Proc. 2014 Dec; 46(10):3515-22.
- 446 38. Bredin C, Naimimohasses S, Norris S, Wright C, Hancock N, Hart K, et al. Development
- and relative validation of a short food frequency questionnaire for assessing dietary intakes of
- 448 non-alcoholic fatty liver disease patients. Eur.J.Nutr. 2019 Feb 25.

| 449 | 39. Cade JE, Burley VJ, Warm DL, Thompson RL, Margetts BM. Food frequency |
|-----|---|
| 450 | questionnaires: A review of their design, validation and utilisation. Nutrition Research Reviews. |
| 451 | 2004;17(1):5-22. |

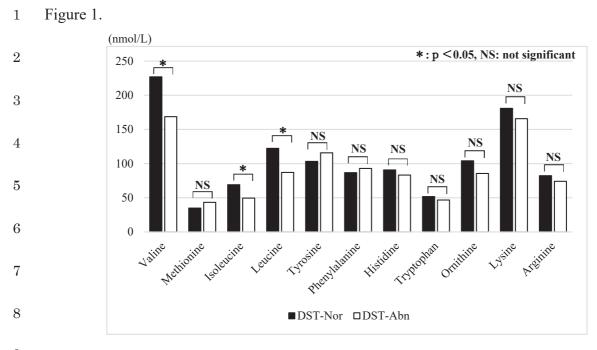
- 452 40. Denissen KFM, Boonen A, Nielen JTH, Feitsma AL, van den Heuvel EGHM, Emans PJ,
- 453 et al. Consumption of dairy products in relation to the presence of clinical knee osteoarthritis:
- 454 The maastricht study. Eur.J.Nutr. 2018 Sep 21.
- 455 41. Eng JY, Moy FM. Validation of a food frequency questionnaire to assess dietary cholesterol,
- 456 total fat and different types of fat intakes among malay adults. Asia Pac.J.Clin.Nutr. 2011 ;
- 457 20(4):639-45.
- 458 42. Kanehara R, Goto A, Kotemori A, Mori N, Nakamura A, Sawada N, et al. Validity and
- 459 reproducibility of a self-administered food frequency questionnaire for the assessment of sugar
- intake in middle-aged japanese adults. Nutrients. 2019 Mar 5; 11(3):10.3390/nu11030554.
- 461 43. Marshall SJ, Livingstone KM, Celis-Morales C, Forster H, Fallaize R, O'Donovan CB, et
- al. Reproducibility of the online Food4Me food-frequency questionnaire for estimating dietary
- 463 intakes across europe. J.Nutr. 2016 May; 146(5):1068-75.
- 464 44. Shim JS, Oh K, Kim HC. Dietary assessment methods in epidemiologic studies.
- 465 Epidemiology and Health. 2014 ; 36:1-8.

| 466 | 45. Peru | ımpa | il BJ, I | Li AA, Cl | nolaı | nkeril | G, Kum | ari 1 | R, Ahmed | A. Optimizi | ng the | nutriti | onal |
|-----|----------|-------|----------|-----------|-------|--------|---------|-------|------------|-------------|--------|---------|------|
| 467 | support | of | adult | patients | in | the | setting | of | cirrhosis. | Nutrients. | 2017 | Oct | 13; |
| 468 | 9(10):10 |).339 | 0/nu91 | 01114. | | | | | | | | | |

- 469 46. Shawcross DL. Is it time to target gut dysbiosis and immune dysfunction in the therapy of
- 470 hepatic encephalopathy? Expert Rev.Gastroenterol.Hepatol. 2015 May; 9(5):539-42.
- 471 47. Malaguarnera G, Giordano M, Nunnari G, Bertino G, Malaguarnera M. Gut microbiota in
- 472 alcoholic liver disease: Pathogenetic role and therapeutic perspectives. World J.Gastroenterol.
- 473 2014 Nov 28; 20(44):16639-48.
- 474 48. Liu Q, Duan ZP, Ha DK, Bengmark S, Kurtovic J, Riordan SM. Synbiotic modulation of
- 475 gut flora: Effect on minimal hepatic encephalopathy in patients with cirrhosis. Hepatology.
- 476 2004 May; 39(5):1441-9.
- 477 49. Zhu S, Zhao J, Chen Z, Wang Y. Influential factors on cognitive performance in middle-
- 478 aged cohort: Third national health and nutrition examination survey-based study. Medicine
- 479 (Baltimore). 2018 Sep; 97(37):e12033.
- 480
- 481

482 Figure legends

- 483 **Figure 1.** The results of the amino acid analysis according to the DST status. Significant
- 484 differences between the two groups were noted in valine (p=0.0016), leucine (p=0.0078), and
- 485 isoleucine (p=0.0022). The tyrosine value was similar between the two groups.



9 DST-Nor, normal digit symbol test; DST-Abn, abnormal digit symbol test.

| Characteristics | All patients (n = 59) Mean ± SD or n (%) | | |
|---|---|--|--|
| | | | |
| | | | |
| Age (years) | 66.1 ± 10.7 | | |
| Sex (M/F) | 36 / 23 | | |
| BMI (kg/m ²) | 24.0 ± 4.7 | | |
| BMI>25 | 19 (32.2) | | |
| Number of cases with reduced muscle volume | 10 (16.9) | | |
| Etiology | | | |
| HBV | 4 (6.8) | | |
| HCV | 24 (40.7) | | |
| Alcohol | 17 (34.7) | | |
| NASH | 6 (10.2) | | |
| others | 8 (13.6) | | |
| Child–Pugh score | 6.8 ± 2.1 | | |
| Child–Pugh classification (A/B/C) | 37 / 14 / 8 | | |
| MELD score | 9.3 ± 4.8 | | |
| Complication | | | |
| HCC | 31 (52.5) | | |
| Ascites | 12 (20.3) | | |
| Number of cases using BCAA supplementation | 27 (45.8) | | |
| BCAA granules, three sachets daily | 13 (22.0) | | |
| BCAA powder mix, one/two/three packages daily | 11 (18.6) /1 (1.7) /2 (3.4) | | |
| Concomitant medications | | | |
| Lactulose | 5 (8.5) | | |
| Antibiotics | 5 (8.5) | | |
| Zinc sulfate | 3 (5.1) | | |
| L-Carnitine | 0 (0.0) | | |
| Molecular-targeted agents for HCC | 0 (0.0) | | |
| <laboratory findings=""></laboratory> | | | |
| ALT (U/L) | 39.1 ± 31.2 | | |
| Albumin (g/dL) | 3.4 ± 0.5 | | |
| Total-bilirubin (mg/dL) | 1.5 ± 1.1 | | |
| Platelet (×10 ⁴ / μ L) | 11.5 ± 5.9 | | |
| Prothrombin time (%) | 73.3 ± 17.0 | | |
| | | | |

1 Table 1. The clinical characteristics and laboratory findings of the enrolled patients

| NH ₃ (µg/dL) | 56.2 ± 27.4 |
|---------------------------------------|------------------|
| CRP (mg/dL) | 0.44 ± 0.64 |
| Amino Acids, standard value | |
| Valine (nmol/L), 158.4 ~ 287.7 | 212.6 ± 57.4 |
| Leucine (nmol/L), 80.9 ~ 154.3 | 113.7 ± 35.5 |
| Isoleucine (nmol/L), 41.3 ~ 84.9 | 64.2 ± 19.3 |
| Methionine (nmol/L), $19.2 \sim 32.7$ | 37.0 ± 14.3 |
| Tyrosine (nmol/L), 50.2 ~ 82.6 | 106.3 ± 31.1 |
| Phenylalanine (nmol/L), 45.7 ~ 76.5 | 88.3 ± 26.6 |
| Histidine (nmol/L), 67.9 ~ 97.1 | 88.9 ± 15.8 |
| Tryptophan (nmol/L), 41.4 ~ 65.5 | 50.5 ± 14.6 |
| Ornithine (nmol/L), 43.2 ~ 95.7 | 99.5 ± 31.2 |
| Lysine (nmol/L), 118.7 ~ 257.0 | 177.1 ± 44.4 |
| Arginine (nmol/L), 46.0 ~ 121.7 | 80.3 ± 28.4 |
| BTR | 3.9 ± 1.6 |

SD, standard deviation; BMI, body mass index; HBV, hepatitis B virus; HCV, hepatitis C virus; NASH, nonalcoholic steatohepatitis; MELD score, model for end-stage liver disease score; HCC, hepatocellular carcinoma; BCAAs, branched-chain amino acids; ALT, alanine transaminase; Na, serum sodium; NH₃, serum ammonia; CRP, C-reactive protein; BTR, branched-chain amino acid to tyrosine ratio.

 $\mathbf{2}$

Table2. The clinical characteristics of patients with normal and abnormal digit symbol testresults

| Characteristics | DST-Nor | DST-Abn | | |
|--|------------------------|------------------------|------------|--|
| Characteristics | Mean \pm SD or n (%) | Mean \pm SD or n (%) | p value | |
| | | | | |
| No. of cases | 45 | 14 | | |
| Age (years) | 66.5 ± 11.3 | 64.6 ± 8.8 | p = 0.52 | |
| Sex (M/F) | 29 / 16 | 7 / 7 | | |
| BMI (kg/m ²) | 23.9 ± 4.8 | 24.5 ± 4.4 | p = 0.65 | |
| Number of cases with reduced muscle volume | 6 (13.3) | 4 (28.6) | p = 0.24 | |
| Child–Pugh score | 6.1 ± 1.3 | 8.8 ± 2.9 | p = 0.0052 | |
| Child–Pugh classification (A/B/C) | 33 / 10 / 2 | 4 / 4 / 6 | | |
| MELD score | 8.6 ± 4.3 | 11.6 ± 5.7 | p = 0.095 | |
| Complication | | | | |
| HCC | 25 (55.6) | 6 (42.9) | p = 0.542 | |
| | | | | |

| Ascites | 6 (13.3) | 6 (42.9) | p = 0.026 |
|---|-----------------------------------|-----------------------------------|-------------|
| Number of cases using BCAA supplementation | 20 (44.4) | 7 (50.0) | p = 0.776 |
| BCAA granules, three sachets daily | 10 (22.2) | 3 (21.4) | |
| BCAA powder mix, one/two/three packages daily | 9 (20.0)/0(0)/0(0) | 2(14.3)/1(7.1)/2(14.3) | |
| Concomitant medications | | | |
| Lactulose | 2 (4.4) | 3 (21.4) | p = 0.081 |
| Antibiotics | 2 (4.4) | 3 (21.4) | p = 0.081 |
| Zinc sulfate | 3 (6.7) | 0 (0.0) | p = 1.00 |
| <laboratory findings=""></laboratory> | ````````````````````````````````` | ````````````````````````````````` | - |
| ALT (U/L) | 36.2 ± 25.1 | 48.3 ± 45.7 | p = 0.36 |
| Albumin (g/dL) | 3.5 ± 0.5 | 3.0 ± 0.5 | p = 0.0043 |
| Total-bilirubin (mg/dL) | 1.2 ± 0.5 | 2.6 ± 1.7 | p = 0.0071 |
| Platelet (×10 ⁴ / μ L) | 11.4 ± 5.5 | 11.9 ± 7.4 | p = 0.80 |
| Prothrombin time (%) | 76.7 ± 14.3 | 62.4 ± 20.9 | p = 0.028 |
| Na (mmol/L) | 140.0 ± 2.6 | 137.6 ± 3.5 | p = 0.035 |
| NH ₃ (µg/dL) | 56.8 ± 28.5 | 54.1 ± 24.7 | p = 0.73 |
| CRP (mg/dL) | 0.4 ± 0.6 | 0.7 ± 0.7 | p = 0.122 |
| Amino Acids, standard value | | | |
| Valine (nmol/L), 158.4 ~ 287.7 | 226.9 ± 49.0 | 168.6 ± 60.9 | p = 0.0016 |
| Leucine (nmol/L), 80.9 ~ 154.3 | 122.3 ± 31.2 | 87.1 ± 36.2 | p = 0.0078 |
| Isoleucine (nmol/L), 41.3 ~ 84.9 | 69.0 ± 17.0 | 49.5 ± 19.1 | p = 0.0022 |
| Methionine (nmol/L), 19.2 ~ 32.7 | 34.9 ± 10.6 | 43.4 ± 21.6 | p = 0.215 |
| Tyrosine (nmol/L), 50.2 ~ 82.6 | 103.3 ± 29.5 | 115.8 ± 35.3 | p = 0.284 |
| Phenylalanine (nmol/L), 45.7 ~ 76.5 | 86.8 ± 22.1 | 93.0 ± 38.2 | p = 0.825 |
| Histidine (nmol/L), 67.9 ~ 97.1 | 90.7 ± 15.4 | 83.2 ± 16.3 | p = 0.175 |
| Tryptophan (nmol/L), 41.4 ~ 65.5 | 51.8 ± 13.2 | 46.8 ± 18.6 | p = 0.364 |
| Ornithine (nmol/L), 43.2 ~ 95.7 | 104.0 ± 32.7 | 85.6 ± 21.9 | p = 0.112 |
| Lysine (nmol/L), 118.7 ~ 257.0 | 180.8 ± 46.9 | 165.7 ± 34.8 | p = 0.376 |
| Arginine (nmol/L), 46.0 ~ 121.7 | 82.3 ± 28.6 | 74.1 ± 28.1 | p = 0.339 |
| BTR | 4.3 ± 1.6 | 2.8 ± 1.1 | p = 0.00025 |

DST-Nor, normal digit symbol test; DST-Abn, abnormal digit symbol test; BMI, body mass index; MELD, model for End-stage Liver Disease; HCC, hepatocellular carcinoma; BCAAs, branched-chain amino acids; ALT, alanine transaminase; Na, serum sodium; NH₃, serum ammonia; C-reactive protein; BTR, branched-chain amino acids to tyrosine ratio.

Table 3. The multivariate analysis of factors associated with a DST abnormality

| parameter | Odds ratio | 95% CI | p value |
|-------------------------|------------|--------------|---------|
| BTR <2.92 | 9.24 | 1.23 - 69.2 | 0.031 |
| Alb <2.90 | 1.63 | 0.175 - 15.2 | 0.669 |
| T-Bil >1.70 | 3.10 | 0.581 - 16.6 | 0.185 |
| PT <70 | 0.772 | 0.101 - 5.90 | 0.803 |
| Na <138 | 3.34 | 0.519 - 21.5 | 0.204 |
| The presence of ascites | 1.13 | 0.100 - 13.0 | 0.923 |

n;

| 11 | DST, | digit symbol | test; 95%CI, | 95% confide | ence interval | ; BTR, | branched | -chain | amino | acids to | o tyrosine | ratio; Alb | , albumin; |
|----|------|--------------|--------------|-------------|---------------|--------|----------|--------|-------|----------|------------|------------|------------|
|----|------|--------------|--------------|-------------|---------------|--------|----------|--------|-------|----------|------------|------------|------------|

T-Bil, total-bilirubin; PT, prothrombin time; Na: serum sodium.

Table 4. Estimation of the usual daily nutritional condition using the FFQ

| | Man (n = 22) | Woman (n = 19) | EAR or AI |
|----------------------------------|------------------|-------------------|-------------------------|
| | Mean±SD | Mean±SD | (man, woman) |
| | | | |
| Age (years) | 68.5 ± 9.1 | 63.5 ± 13.3 | |
| Height (cm) | 166.0 ± 8.5 | 154.4 ± 4.1 | |
| Body weight (kg) | 63.9 ± 14.3 | 58.6 ± 11.9 | |
| Ideal body weight (kg) | 60.8 ± 6.1 | 52.5 ± 2.8 | |
| BMI | 23.0 ± 3.9 | 24.7 ± 5.6 | |
| TEE (kcal) | 1769.4 ± 309.3 * | 1515.7 ± 149.7 ** | |
| TEE / body weight (kcal/kg) | 26.5 ± 1.6 | 28.1 ± 3.4 | |
| <results ffq="" of=""></results> | | | |
| EIE (kcal/day) | 1932.7 ± 533.4 * | 1636.9 ± 455.8 ** | 2200-2650, 1700-2000 |
| Ratio of EIE to EER | 0.94 ± 0.29 | 0.93 ± 0.29 | |
| EIE / body weight (kcal/kg) | 30.9 ± 8.0 | 28.9 ± 8.5 | |
| TEE / EIE | 1.0 ± 0.5 | 1.0 ± 0.3 | |
| Protein intake (g/kg) | 1.2 ± 0.3 | 1.1 ± 0.4 | |

| <usual daily="" energy="" ratio=""></usual> | | | |
|---|----------------|---------------|------------|
| Protein (%) | 15.1 ± 1.8 | 14.8 ± 2.5 | 20-30 |
| Fat (%) | 28.6 ± 5.2 | 30.2 ± 4.5 | 20-25 |
| Carbohydrates (%) | 56.6 ± 5.7 | 54.9 ± 6.2 | 50-70 |
| Dietary fiber (g/day) | 16.2 ± 6.0 | 16.2 ± 4.4 | 19<, 17< |
| n-3 USFA (g/day) | 2.3 ± 1.0 | 1.8 ± 1.7 | 2.2<, 1.8< |
| n-6 USFA (g/day) | 9.6 ± 3.9 | 8.9 ± 3.5 | 8-10, 7-9 |
| n-6 USFA/n-3 USFA | 4.5 ± 1.6 | 5.1 ± 1.4 | |
| Salt (g/day) | 9.8 ± 2.8 | 8.9 ± 2.9 | 9>, 7.5> |

SD, standard deviation; TEE, total daily energy expenditure; EIE, estimated intake energy per day;

EER, estimated energy requirement; EAR, estimated average requirement;

n-3 USFA, n-3 unsaturated fatty acid; n-6 USFA, n-6 unsaturated fatty acid; AI, adequate intake;

*/**, There was no significant difference for the comparison of TEE and EIE; based on sex.

20

Table 5. The nutritional findings of patients with normal and abnormal digit symbol test results

| | DST-Nor | DST-Abn | |
|---|--------------------|--------------------|---------|
| Parameter | (n = 34) | (n = 7) | p value |
| | Mean±SD | Mean±SD | |
| | | | |
| Sex (Man:Wonam) | 19:15 | 2:5 | |
| < <results ffq="" of="">></results> | | | |
| EIE (kcal/day) | 1814.4 ± 553.7 | 1704.7 ± 136.4 | p=0.826 |
| Ratio of EIE to EER | 0.94 ± 0.31 | 0.94 ± 0.16 | p=0.852 |
| EIE / body weight (kcal/kg) | 29.8 ± 8.3 | 33.0 ± 8.2 | p=0.466 |
| Protein intake (g/kg) | 1.1 ± 0.4 | 1.2 ± 0.4 | p=0.834 |
| <usual daily="" energy="" ratio=""></usual> | | | |
| Protein (%) | 15.2 ± 2.0 | 13.8 ± 2.1 | p=0.141 |
| Fat (%) | 29.3 ± 5.0 | 28.6 ± 4.2 | p=0.533 |
| Carbohydrates (%) | 55.5 ± 6.1 | 57.7 ± 4.3 | p=0.200 |
| Dietary fiber (g/day) | 16.5 ± 5.4 | 14.6 ± 3.2 | p=0.703 |
| n-3 USFA (g/day) | 2.1 ± 0.9 | 1.9 ± 0.4 | p=0.599 |
| n-6 USFA (g/day) | 9.4 ± 3.9 | 8.8 ± 2.2 | p=0.795 |
| | | | |

| n-6 USFA/n-3 USFA | 4.8 ± 1.5 | 4.9 ± 1.5 | p=0.876 |
|-------------------|-------------|-------------|---------|
| Salt (g/day) | 9.6 ± 2.9 | 8.4 ± 1.7 | p=0.267 |

DST-Nor, normal digit symbol test; DST-Abn, abnormal digit symbol test;

EIE, estimated intake energy per day; EER, estimated energy requirements;

n-3 USFA, n-3 unsaturated fatty acid; n-6 USFA, n-6 unsaturated fatty acid.

23

24 Table 6. The clinical characteristics of LC patients that do not take BCAA supplementation

25 with normal and abnormal digit symbol test results

| Mean \pm SD or n (%)Mean \pm SD or n (%)Number of cases257Age (years) 66.9 ± 10.9 61.7 ± 10.1 $p = 0.266$ bian (MF) $18/7$ $6/1$ SMI (kg/m ²) 24.2 ± 4.9 24.4 ± 3.4 $p = 0.915$ Sumber of cases with reduced muscle volume 1 (4.0) 2 (28.6) $p = 0.12$ Child-Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child-Pugh classification (A/B/C) $21/3/1$ $3/1/3$ $71/3$ AELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication $16C$ 18 (72.0) 2 (28.6) $p = 0.074$ Ascites 5 (20.0) 3 (42.9) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) $p = 0.327$ Concomitant medications 14.0 0 (0.0) $p = 0.327$ Lactulose 0 (0.0) 0 (0.0) p | Characteristics | DST-Nor | DST-Abn | p value | |
|--|---------------------------------------|------------------------|------------------------|----------------|--|
| Age (years) 66.9 ± 10.9 61.7 ± 10.1 $p = 0.266$ Sex (M/F) $18/7$ $6/1$ SMI (kg/m ²) 24.2 ± 4.9 24.4 ± 3.4 $p = 0.915$ Sumber of cases with reduced muscle volume $1 (4.0)$ $2 (28.6)$ $p = 0.12$ Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21/3/1$ $3/1/3$ $71/3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $2 (28.6)$ $p = 0.074$ Lactulose $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ Latulose $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ Paboratory findings> 3.6 ± 25.1 46.6 ± 58.1 NLT (U/L) 3.6 ± 0.5 3.0 ± 0.6 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.651 1.0 ± 0.5 2.8 ± 2.0 Contarbilizubin (mg/LL) 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.652 3.0 ± 0.6 $p = 0.326$ Ada (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 Pa = 0.5251 46.6 ± 58.1 | Characteristics | Mean \pm SD or n (%) | Mean \pm SD or n (%) | p value | |
| Age (years) 66.9 ± 10.9 61.7 ± 10.1 $p = 0.266$ Sex (M/F) $18/7$ $6/1$ SMI (kg/m ²) 24.2 ± 4.9 24.4 ± 3.4 $p = 0.915$ Sumber of cases with reduced muscle volume $1 (4.0)$ $2 (28.6)$ $p = 0.12$ Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21/3/1$ $3/1/3$ $71/3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $2 (28.6)$ $p = 0.074$ Lactulose $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ Latulose $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ Paboratory findings> 3.6 ± 25.1 46.6 ± 58.1 NLT (U/L) 3.6 ± 0.5 3.0 ± 0.6 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.641 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.651 1.0 ± 0.5 2.8 ± 2.0 Contarbilizubin (mg/LL) 1.0 ± 0.5 2.8 ± 2.0 Pa = 0.652 3.0 ± 0.6 $p = 0.326$ Ada (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 Pa = 0.5251 46.6 ± 58.1 | | | _ | | |
| iex (M/F) $18/7$ $6/1$ BMI (kg/m ²) 24.2 ± 4.9 24.4 ± 3.4 $p = 0.915$ Sumber of cases with reduced muscle volume 1 (4.0) 2 (28.6) $p = 0.12$ Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21/3/1$ $3/1/3$ 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication RCC 18 (72.0) 2 (28.6) $p = 0.074$ Ascites 5 (20.0) 3 (42.9) $p = 0.327$ Concomitant medications R $0 = 0.00$ Lactulose 0 (0.0) 0 (0.0) $p = 0.641$ Athibiotics 0 (0.0) 0 (0.0) $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 | | | | 0.044 | |
| BMI (kg/m^2) 24.2 ± 4.9 24.4 ± 3.4 $p = 0.915$ Sumber of cases with reduced muscle volume $1 (4.0)$ $2 (28.6)$ $p = 0.12$ Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21 / 3 / 1$ $3 / 1 / 3$ $3 / 1 / 3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication HCC $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $Lactulose$ $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ $p = 0.641$ Altropy findings> $XLT (U/L)$ 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.6251$ Chatelet ($\times 10^4/\mu L$) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.2251$ Prothrombin time (%) 80.3 ± 13.3 69.4 | | | | p = 0.266 | |
| Number of cases with reduced muscle volume1 (4.0)2 (28.6) $p = 0.12$ Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21 / 3 / 1$ $3 / 1 / 3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication PCC $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $Lactulose$ $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ $p = 1.00$ Aboratory findings> $XLT (U/L)$ 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Patelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.121$ NH3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | | | | 0 0 1 - | |
| Child—Pugh score 5.9 ± 1.3 8.4 ± 3.6 $p = 0.121$ Child—Pugh classification (A/B/C) $21 / 3 / 1$ $3 / 1 / 3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication HCC $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $Lactulose$ $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ $p = 1.00$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ $p = 0.641$ Albumin (g/dL) 3.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ NH _3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | | | | 1 | |
| Child—Pugh classification (A/B/C) $21 / 3 / 1$ $3 / 1 / 3$ MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication18 (72.0) 2 (28.6) $p = 0.074$ Ascites 5 (20.0) 3 (42.9) $p = 0.327$ Concomitant medications 2 (0.0) 0 (0.0) 0 (0.0)Lactulose 0 (0.0) 0 (0.0) $p = 1.00$ Zinc sulfate 1 (4.0) 0 (0.0) $p = 1.00$ Claboratory findings> 3.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ | | | | - | |
| MELD score 7.6 ± 4.8 9.9 ± 7.3 $p = 0.469$ Complication18 (72.0)2 (28.6) $p = 0.074$ Ascites5 (20.0)3 (42.9) $p = 0.327$ Concomitant medications $2 (28.6)$ $p = 0.327$ Lactulose0 (0.0)0 (0.0)Antibiotics0 (0.0)0 (0.0)Zinc sulfate1 (4.0)0 (0.0)Caboratory findings> 3.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Abumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.822$ | | 5.9 ± 1.3 | 8.4 ± 3.6 | p = 0.121 | |
| TomplicationHCC18 (72.0)2 (28.6) $p = 0.074$ Ascites5 (20.0)3 (42.9) $p = 0.327$ Concomitant medications $1 (4.0)$ 0 (0.0) $0 (0.0)$ Antibiotics0 (0.0)0 (0.0) $0 (0.0)$ Zinc sulfate1 (4.0)0 (0.0) $p = 1.00$ Caboratory findings> 3.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Total-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Child–Pugh classification (A/B/C) | 21 / 3 / 1 | 3 / 1 / 3 | | |
| HCC $18 (72.0)$ $2 (28.6)$ $p = 0.074$ Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $2 (0.0)$ $3 (42.9)$ $p = 0.327$ Lactulose $0 (0.0)$ $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ $p = 1.00$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ $p = 1.00$ ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Total-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | MELD score | 7.6 ± 4.8 | 9.9 ± 7.3 | p = 0.469 | |
| Ascites $5 (20.0)$ $3 (42.9)$ $p = 0.327$ Concomitant medications $0 (0.0)$ $0 (0.0)$ $0 (0.0)$ Lactulose $0 (0.0)$ $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ $p = 1.00$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ $p = 1.00$ Claboratory findings> 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Total-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ H3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Complication | | | | |
| Concomitant medicationsLactulose $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ Claboratory findings>ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 H3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 | НСС | 18 (72.0) | 2 (28.6) | p = 0.074 | |
| Lactulose $0 (0.0)$ $0 (0.0)$ Antibiotics $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ CLaboratory findings>ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 | Ascites | 5 (20.0) | 3 (42.9) | p = 0.327 | |
| Antibiotics $0 (0.0)$ $0 (0.0)$ Zinc sulfate $1 (4.0)$ $0 (0.0)$ $p = 1.00$ Claboratory findings>ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Concomitant medications | | | | |
| Zinc sulfate1 (4.0)0 (0.0) $p = 1.00$ CLaboratory findings>ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Va (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Lactulose | 0 (0.0) | 0 (0.0) | | |
| CLaboratory findings>ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Va (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Antibiotics | 0 (0.0) | 0 (0.0) | | |
| ALT (U/L) 35.6 ± 25.1 46.6 ± 58.1 $p = 0.641$ Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Va (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Zinc sulfate | 1 (4.0) | 0 (0.0) | p = 1.00 | |
| Albumin (g/dL) 3.6 ± 0.5 3.0 ± 0.6 $p = 0.05$ Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | <laboratory findings=""></laboratory> | | | | |
| Cotal-bilirubin (mg/dL) 1.0 ± 0.5 2.8 ± 2.0 $p = 0.06$ Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | ALT (U/L) | 35.6 ± 25.1 | 46.6 ± 58.1 | p = 0.641 | |
| Platelet (×10 ⁴ /µL) 11.8 ± 5.9 15.9 ± 8.1 $p = 0.251$ Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Na (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ NH ₃ (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Albumin (g/dL) | 3.6 ± 0.5 | 3.0 ± 0.6 | p = 0.05 | |
| Prothrombin time (%) 80.3 ± 13.3 69.4 ± 26.3 $p = 0.326$ Ma (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ MH3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Total-bilirubin (mg/dL) | 1.0 ± 0.5 | 2.8 ± 2.0 | p = 0.06 | |
| Ja (mmol/L) 140.2 ± 2.1 137.0 ± 4.7 $p = 0.125$ JH3 (µg/dL) 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Platelet (×10 ⁴ / μ L) | 11.8 ± 5.9 | 15.9 ± 8.1 | p = 0.251 | |
| $MH_3 \ (\mu g/dL)$ 52.3 ± 25.2 50.9 ± 16.6 $p = 0.862$ | Prothrombin time (%) | 80.3 ± 13.3 | 69.4 ± 26.3 | p = 0.326 | |
| | Na (mmol/L) | 140.2 ± 2.1 | 137.0 ± 4.7 | p = 0.125 | |
| CRP (mg/dL) 0.31 ± 0.37 0.92 ± 0.77 $p = 0.08$ | NH ₃ (µg/dL) | 52.3 ± 25.2 | 50.9 ± 16.6 | p = 0.862 | |
| | CRP (mg/dL) | 0.31 ± 0.37 | 0.92 ± 0.77 | p = 0.08 | |

| Valine (nmol/L), 158.4 ~ 287.7 | 222.6 ± 42.6 | 169.7 ± 77.6 | p = 0.127 |
|-------------------------------------|-----------------|-----------------|-------------|
| Leucine (nmol/L), 80.9 ~ 154.3 | 119.5 ± 27.9 | 93.5 ± 45.9 | p = 0.195 |
| Isoleucine (nmol/L), 41.3 ~ 84.9 | 69.4 ± 14.8 | 50.6 ± 24.3 | p = 0.09 |
| Methionine (nmol/L), 19.2 ~ 32.7 | 32.6 ± 9.3 | 29.2 ± 8.8 | p = 0.403 |
| Tyrosine (nmol/L), 50.2 ~ 82.6 | 100.2 ± 33.0 | 97.5 ± 26.0 | p = 0.824 |
| Phenylalanine (nmol/L), 45.7 ~ 76.5 | 84.6 ± 22.9 | 74.4 ± 21.0 | p = 0.288 |
| Histidine (nmol/L), 67.9 ~ 97.1 | 90.0 ± 13.3 | 73.8 ± 11.0 | p = 0.00691 |
| Tryptophan (nmol/L), 41.4 ~ 65.5 | 52.4 ± 15.5 | 43.0 ± 15.3 | p = 0.185 |
| Ornithine (nmol/L), 43.2 ~ 95.7 | 105.1 ± 29.9 | 81.3 ± 21.8 | p = 0.036 |
| Lysine (nmol/L), 118.7 ~ 257.0 | 188.7 ± 49.9 | 158.4 ± 39.3 | p = 0.117 |
| Arginine (nmol/L), 46.0 ~ 121.7 | 83.1 ± 26.2 | 57.2 ± 19.0 | p = 0.012 |
| BTR | 4.58 ± 1.83 | 3.24 ± 1.21 | p = 0.0385 |

DST-Nor, normal digit symbol test; DST-Abn, abnormal digit symbol test; BMI, body mass index; MELD, model for End-stage Liver Disease; HCC, hepatocellular carcinoma; BCAAs, branched-chain amino acids; ALT, alanine transaminase; Na, serum sodium; NH₃, serum ammonia; C-reactive protein; BTR, branched-chain amino acids to tyrosine ratio.