



# A combination of desmopressin and docetaxel inhibit cell proliferation and invasion mediated by urokinase-type plasminogen activator (uPA) in human prostate cancer cells



Hiroshi Sasaki <sup>a</sup>, Laurence H. Klotz <sup>a</sup>, Linda M. Sugar <sup>b</sup>, Alexander Kiss <sup>c</sup>,  
Vasundara Venkateswaran <sup>a,\*</sup>

<sup>a</sup> Division of Urology, Sunnybrook Health Sciences Center, Toronto, ON, Canada

<sup>b</sup> Department of Pathology, Sunnybrook Health Sciences Center, Toronto, ON, Canada

<sup>c</sup> Department of Research Design and Biostatistics, Institute for Clinical Evaluative Sciences, Sunnybrook Health Sciences Center, Toronto, ON, Canada

## ARTICLE INFO

### Article history:

Received 5 July 2015

Accepted 9 July 2015

Available online 13 July 2015

### Keywords:

Prostate cancer

Desmopressin

Docetaxel

uPA

Matrix metalloproteinase

## ABSTRACT

**Background:** This study was designed to assess the effectiveness of a combination treatment using both desmopressin and docetaxel in prostate cancer treatment. Desmopressin is a well-known synthetic analogue of the antidiuretic hormone vasopressin. It has recently been demonstrated to inhibit tumor progression and metastasis in *in vivo* models. Docetaxel is widely used for the treatment of castration resistant prostate cancer (CRPC) patients. However, durable responses have been uncommon to date. In this study, we investigated the anti-tumor effect of desmopressin in combination with docetaxel *in vitro* and *in vivo*.

**Methods:** Two prostate cancer cells (PC3, LNCaP) were treated with different concentrations of desmopressin alone, docetaxel alone, and a combination of desmopressin and docetaxel. Cell proliferation was determined by MTS assay. The anti-invasive and anti-migration potential of desmopressin and in combination with docetaxel were examined by wound healing assay, migration chamber assay, and matrigel invasion assay.

**Results:** The combination of desmopressin and docetaxel resulted in a significant inhibition of PC3 and LNCaP cell proliferation ( $p < 0.01$ ). Additionally, cell migration and invasion were also inhibited by the combination when compared to that of either treatment alone in PC3 cells ( $p < 0.01$ ). The anti-tumor effect of this combination treatment was associated with down-regulation of both urokinase-type plasminogen activator (uPA) and matrix metalloproteinase (MMP-2 and MMP-9) in PC3 cells.

**Conclusions:** We are the first to elucidate the anti-tumor and anti-metastatic potential of desmopressin in combination with docetaxel in a prostate cancer model via the uPA-MMP pathway. Our finding could potentially contribute to the therapeutic profile of desmopressin and enhance the efficacy of docetaxel based treatment for CRPC.

© 2015 Elsevier Inc. All rights reserved.

## 1. Introduction

Docetaxel is the first cytotoxic drug of choice for management of CRPC [1,2]. Although docetaxel-based combination chemotherapy has significantly improved survival of CRPC patients, durable responses are uncommon [2,3]. Furthermore, docetaxel causes

\* Corresponding author. Department of Surgery, University of Toronto Division of Urology, Sunnybrook Health Sciences Centre, 2075 Bayview Avenue, Toronto, Ontario, M4N 3M5, Canada.

E-mail address: [vasundara.venkateswaran@sunnybrook.ca](mailto:vasundara.venkateswaran@sunnybrook.ca) (V. Venkateswaran).

adverse events such as grade 3 or 4 neutropenia, fatigue, alopecia, and nausea. High doses of docetaxel induce significant toxicity [2,3]. A non-toxic agent that enhances the efficacy of docetaxel without increasing toxicity would enhance the treatment of CRPC. Therefore, a lower dose of docetaxel could be achieved in a combination treatment, and potentially provide us with a better outcome. Desmopressin is a synthetic derivative of the antidiuretic hormone vasopressin. Desmopressin is a safe and effective hemostatic agent in patients with von Willebrand disease, hemophilia A, and other bleeding disorders [4,5]. Recent reports suggest that desmopressin inhibits tumor progression and metastasis in *in vivo*

models [6–8]. Although these studies suggest that desmopressin has both anti-tumor and anti-metastatic effects on cancer cells, the mechanism of action has not been determined.

The aim of this study was to investigate the anti-tumor effects of desmopressin in combination with the chemotherapeutic agent docetaxel *in vitro* and *in vivo*. We demonstrate that desmopressin monotherapy in cultured prostate cancer cells resulted in impeding proliferation. Furthermore, we have delineated a potential mechanism of action of desmopressin on human prostate cancer cells, whereby treatment significantly decreased uPA, MMP-2 and MMP-9 expression levels in PC3 cells.

## 2. Materials and methods

### 2.1. Cell culture

Two human prostate cancer cell lines (PC3 and LNCaP) were obtained from the American Type Culture Collection (Rockville, MD, USA). All cells were maintained as previously described [9,10].

### 2.2. Chemicals

Docetaxel was purchased from Santa Cruz Biotechnology (CA, USA). Docetaxel was prepared in dimethyl sulfoxide (DMSO; Sigma–Aldrich, MO, USA) and diluted with a cell culture medium at a final concentration of 0.01% DMSO. Desmopressin was provided by Ferring Inc (ON, Canada).

### 2.3. Cell proliferation assay

Cell proliferation was determined by MTS assay as previously described [9,10]. Cells ( $5 \times 10^3$ /well) were plated using 96 well plates. After 24 h of cultivation, a range of concentrations of desmopressin (0 nM–1  $\mu$ M) and/or docetaxel (0 nM–100 nM) were added and the culture was allowed to continue for up to 72 h.

### 2.4. Western blot analysis

Protein lysates from the desmopressin monotherapy dose response study (1 nM–1  $\mu$ M), combination study and concentrated media employed for Western blot analysis, were prepared as previously described [9–11]. The membranes were incubated with primary antibodies against Bax, bcl-2, p21, p27, cdk2, uPA, uPAR, MMP-2 and MMP-9 (1:100–200, Santacruz Biotechnology, CA, USA).

### 2.5. Flow cytometry

To analyze cell cycle profiles, cells were plated at a density of  $1 \times 10^6$  per 10 cm dish. Asynchronously growing cells were pulse labeled with 10 mM bromodeoxyuridine (BrdU) for 2 h with or without prior treatment of the antioxidants at the end of 24 h. Cell cycle analysis was carried out on the FACS Calibur flow cytometer using the Cell Quest Pro software package (Becton–Dickinson, CA, USA). Ten thousand events were counted for each experiment.

### 2.6. Wound healing assay

Cell motility was assessed using a wound healing assay, performed according to a protocol described by Liang et al. [12]. Cells ( $2 \times 10^4$ /well) were plated in 6-well plates. The cells were treated with desmopressin and/or docetaxel and allowed to migrate for 24 h. A computer-based microscopy imaging system with a microscope at 200X magnification was used to determine wound healing after scratching the plate.

### 2.7. Cell migration and invasion assay

The migration of cells was measured using transwell insert plates (BD Biosciences, MA, USA) according to the manufacture's protocol. Cells were subjected to 24 h of serum deprivation in DMEM/F12 supplemented with FBS. PC3 ( $5 \times 10^4$ ) cells were plated onto filters in 8.0  $\mu$ m transwell insert plates and treated with these compounds - desmopressin (1 nM, 1  $\mu$ M) and 5 nM docetaxel, in serum-free medium. Bottom wells were filled with complete medium. Cells were allowed to migrate for 24 h. After the treatment, cells remaining on the top surface of the filter were removed using cotton swabs. The migrated cells from random fields were chosen then counted, using the computer-based microscopy imaging system. For the invasion assay, the same procedures were performed as described in the migration assay, except that the cells were plated onto 24-well matrigel-coated transwell plates (BD Biosciences, MA, USA).

### 2.8. Elisa activity assay

PC3 cells were seeded in a six-well plate ( $1.0 \times 10^5$  per well) and incubated at 37 °C. After 24 h, the complete medium was removed and cells were washed with serum-free medium. Cells were incubated in serum-free medium supplemented with desmopressin (1 nM and 1  $\mu$ M) and/or 5 nM docetaxel for 24 h. MMP-9, MMP-2 and uPA activity in the cell lysate was quantified using human MMP-9, MMP-2 and uPA assay kit (R&D Systems, MN, USA) according to the manufacturer's instruction.

### 2.9. In vivo studies using xenografts

The mice were housed and maintained in laminar flow cabinets, under specific pathogen-free conditions, in facilities approved by the University of Toronto Animal Research Ethics Board, and in accordance with their regulations and standards by the Canadian Council on Animal Care (CCAC). Cells ( $1 \times 10^6$  PC3 cells with 100  $\mu$ l martrigel solution (BD Biosciences, CA, USA)) were inoculated subcutaneously (sc) into 6–8 week-old male nude mice (Charles river, QC, CANADA). After 14 days, the developing tumors were measured and mice randomly assigned to different treatment groups. Tumor volumes were determined by measurement of tumor length (L) and width (W) with a caliper and calculated twice a week according to the following formula:  $V = (L \times W^2) (\pi/6)$ . Mice were randomized into four groups; control (n = 15), Desmopressin alone (n = 15), docetaxel alone (n = 10) and desmopressin in combination with docetaxel (n = 10). Mice received desmopressin intravenously in the saline at a final dose of 2  $\mu$ g/ml/body weight (50 ng/0.3 ml saline dose). Mice were administered docetaxel at a dose of 5 mg/kg intravenously as every 3 weeks. Desmopressin was administered in 2 doses, 30 min prior to and 24 h after the administration of docetaxel. The animals that were administered docetaxel or desmopressin in combination with docetaxel were sacrificed 35 days after cell inoculation.

### 2.10. Statistical analyses

Each experiment was repeated 2–3 times and the corresponding statistical analysis was performed. The data represented mean  $\pm$  the standard error of the mean. Statistical analysis was done by Student's t test at a significance level of  $p < 0.05$  using SAS Version 9.3 (SAS Institute, Cary, NC, USA). Analyses of the *in vivo* results were performed using either the Student's t-test, or repeated measures of one-way ANOVA techniques.

### 3. Results

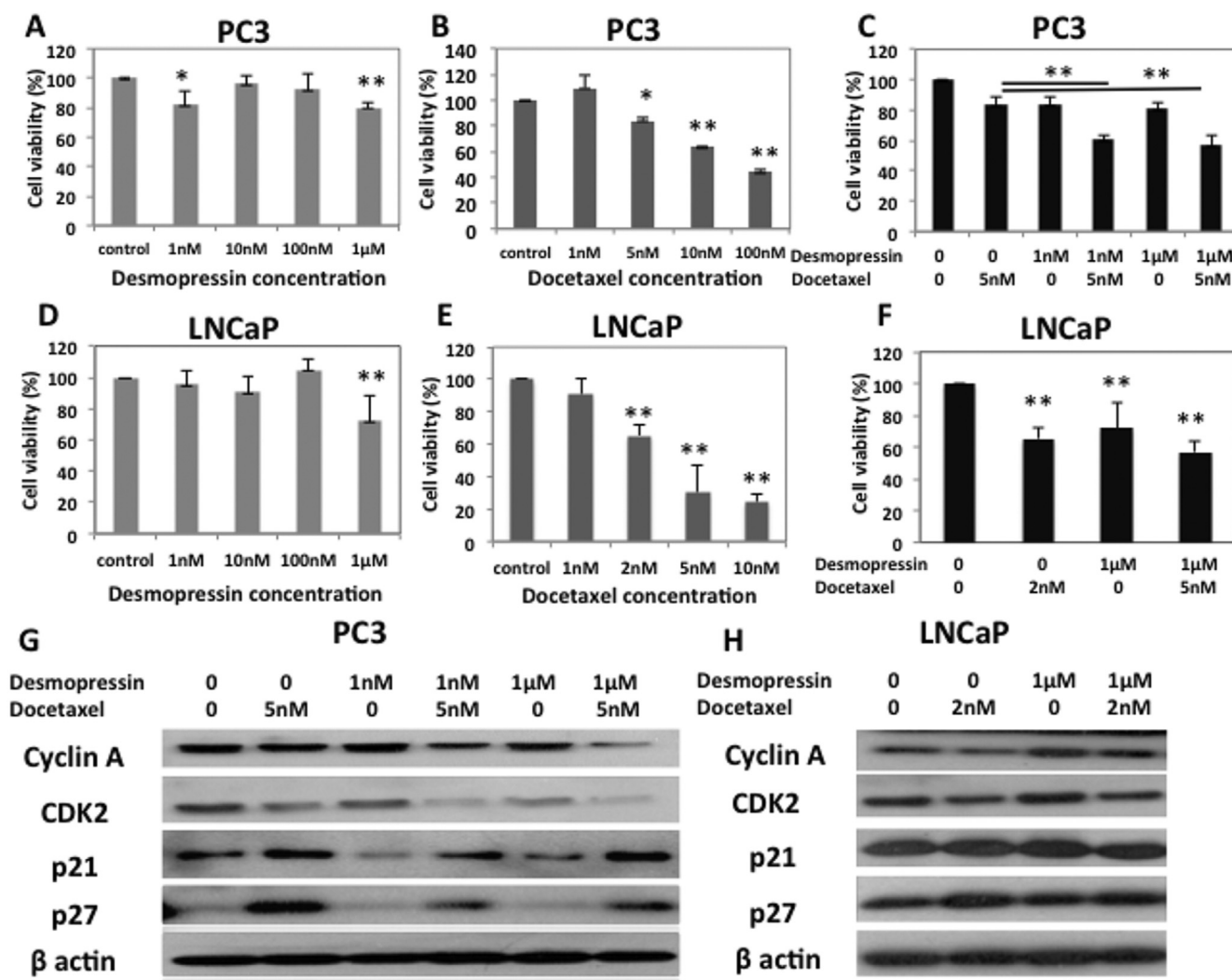
#### 3.1. Docetaxel and/or desmopressin inhibit cell proliferation in prostate cancer cells

The MTS cell proliferation assay was carried out at 72 h with the cells treated with a range of doses, 0 nM–1  $\mu$ M desmopressin and 0 nM–100 nM docetaxel. PC3 cells, treated with 1 nM and 1  $\mu$ M of desmopressin, resulted in an 18% and 20% reduction in cell proliferation (Fig. 1, A, \* $p$  < 0.05 and \*\* $p$  < 0.01). Treatment of PC3 cells with docetaxel for 72 h decreased cell viability in a dose dependent manner (Fig. 1, B). In PC3 cells, combination therapy of desmopressin (1 nM, 1  $\mu$ M) and 5 nM docetaxel resulted in a significant decrease in cell proliferation when compared to treatment with desmopressin as monotherapy (Fig. 1, C, \*\* $p$  < 0.01). LNCaP cells treated with 1  $\mu$ M of desmopressin resulted in a 32% reduction in cell proliferation (Fig. 1, D,

\*\* $p$  < 0.01). Also, treatment of LNCaP cells with docetaxel for 72 h decreased cell viability in a dose dependent manner (Fig. 1, E). In LNCaP cells, combination therapy of desmopressin (1  $\mu$ M) and 2 nM docetaxel resulted in a significant decrease in cell proliferation when compared to treatment with desmopressin as monotherapy (Fig. 1, F, \*\* $p$  < 0.01).

#### 3.2. Desmopressin enhances the apoptotic effect of docetaxel as determined by western blot analysis, and desmopressin in combination with docetaxel induces cell cycle arrest

Docetaxel has the ability to alter key regulatory molecules, including the suppression of microtubules with consequent mitotic spindle disruption, leading to G2/M phase cell cycle arrest and induction of bcl-2 phosphorylation ultimately leading to apoptosis [13,14]. In PC3 cells, the expression of cyclin A and CDK2 were reduced in the cells treated with desmopressin and docetaxel



**Fig. 1.** Cell proliferation assay after 72 h. (A) MTS assay with PC3 cells treated with a dose range of 0–1  $\mu$ M desmopressin for 72 h. (B) MTS assay with PC3 cells treated with a dose range of 0–100 nM docetaxel for 72 h. Significant reductions were seen after 5 nM. (C) PC3 cells were incubated in the presence of the indicated concentration of each drug for 72 h. Each combination therapy resulted in a significant reduction in proliferation when compared to control and corresponding docetaxel alone as well as desmopressin alone groups (\* $p$  < 0.05 and \*\* $p$  < 0.01, Student's t-test). (D) MTS assay with LNCaP cells treated with a dose range of 0–1  $\mu$ M desmopressin for 72 h. (E) MTS assay with LNCaP cells treated with a dose range of 0–10 nM docetaxel for 72 h. Significant reductions were seen after 2 nM. (F) LNCaP cells were incubated in the presence of the indicated concentration of each drug for 72 h. The combination therapy resulted in a significant reduction in proliferation when compared to the control and corresponding docetaxel alone, as well as desmopressin alone group (\*\* $p$  < 0.01, Student's t-test). (G) Western blot shows the effect of a combination treatment of desmopressin and docetaxel on cell cycle regulatory proteins in PC3 cells. (H) Western blot shows the effect of a combination treatment of desmopressin and docetaxel on cell cycle regulatory proteins in LNCaP cells.

(Fig. 1, G). In addition, the expression of CDK inhibitory proteins, p21(waf1/cip1) and p27(kip1), were elevated under the same conditions (Fig. 1, G). These data suggest that desmopressin in combination with docetaxel therapy inhibit the molecules associated with cell cycle progression, and at the same time induce cell cycle arrest in PC3 cells. However, in LNCaP cells p21 and p27 were not significant elevated under the treatment with desmopressin and desmopressin (Fig. 1, H). In terms of the apoptotic pathway, docetaxel monotherapy significantly inhibited bcl-2 expression in both cell lines (Supplementary Fig. 1, A, B). The ratio of Bax/Bcl-2 revealed a 4-fold and 8-fold increased expression in cells treated with 1 nM desmopressin in combination with docetaxel and 1  $\mu$ M desmopressin in combination with docetaxel respectively in PC3 cells (Supplementary Fig. 1, C). However in LNCaP cells, the ratio of Bax/Bcl-2 revealed a one-third decreased expression in cells treated with 1  $\mu$ M desmopressin in combination with docetaxel. This result indicates that desmopressin may influence the apoptotic effect of docetaxel in PC3 cells.

### 3.3. Desmopressin does not alter cell cycle distribution on PC3 cells

We investigated alterations in cell cycle profiles by flow cytometric analysis using BrdU labeling on PC3 cells treated with desmopressin and docetaxel alone and in combination. Treatment with 5 nM docetaxel or a combination with 1 nM/1  $\mu$ M desmopressin showed a significant increase in the proportion of cells in G2 phase, consistent with a G2M cell cycle arrest (Supplementary Fig. 1, D). It should be noted that there was no change in cell cycle distribution following treatment of cells with desmopressin monotherapy.

### 3.4. Differential influences of desmopressin on migration of PC3 cells in a dose dependent manner

Since desmopressin and docetaxel treatment resulted in an anti-proliferative effect on PC3 cells and LNCaP cells in culture, we examined the effect of desmopressin alone and in combination with docetaxel treatment on cell migration with the wound-healing assay. As shown in Fig. 2, the difference in distance migrated by the control between 0 h and 24 h was measured and compared with that of the treated PC3 cells. At the end of 24 h, there was significant decreased cell migration with desmopressin monotherapy compared to the control ( $p < 0.05$ ). We noticed a significant decrease in cell migration with a combination of 1 nM and 1  $\mu$ M desmopressin and 5 nM docetaxel than with either treatment alone on PC3 (Fig. 2, A). However, we could not demonstrate anti-migrative effect during 72h on LNCaP (data not shown). The migratory capability of PC3 cells was further quantified using the migration chamber transwell plates. Different doses of desmopressin (1 nM and 1  $\mu$ M) or docetaxel monotherapy treatment significantly inhibited cell migration (Fig. 2). Moreover, each combination treatment with desmopressin (1 nM and 1  $\mu$ M) and 5 nM docetaxel resulted in a further significant reduction in cell migration compared to controls and either monotherapy (Fig. 2, B and C,  $^{**}p < 0.01$ ).

### 3.5. Desmopressin exhibits anti-invasive potential evidenced by its effect on PC3 cells

To assess the anti-metastatic ability of desmopressin, we performed a matrigel invasion assay. As shown in Fig. 3, 5 nM docetaxel alone and the two different doses of desmopressin (1 nM and 1  $\mu$ M) treatment induced significant inhibition of cell invasion. Furthermore, each combination treatment of 5 nM docetaxel and desmopressin (1 nM and 1  $\mu$ M) resulted in a significant reduction in

cell invasion when compared to that of the control and either monotherapy (Fig. 2, D and E,  $^{**}p < 0.01$ ), indicative of the anti-invasive ability of this combination.

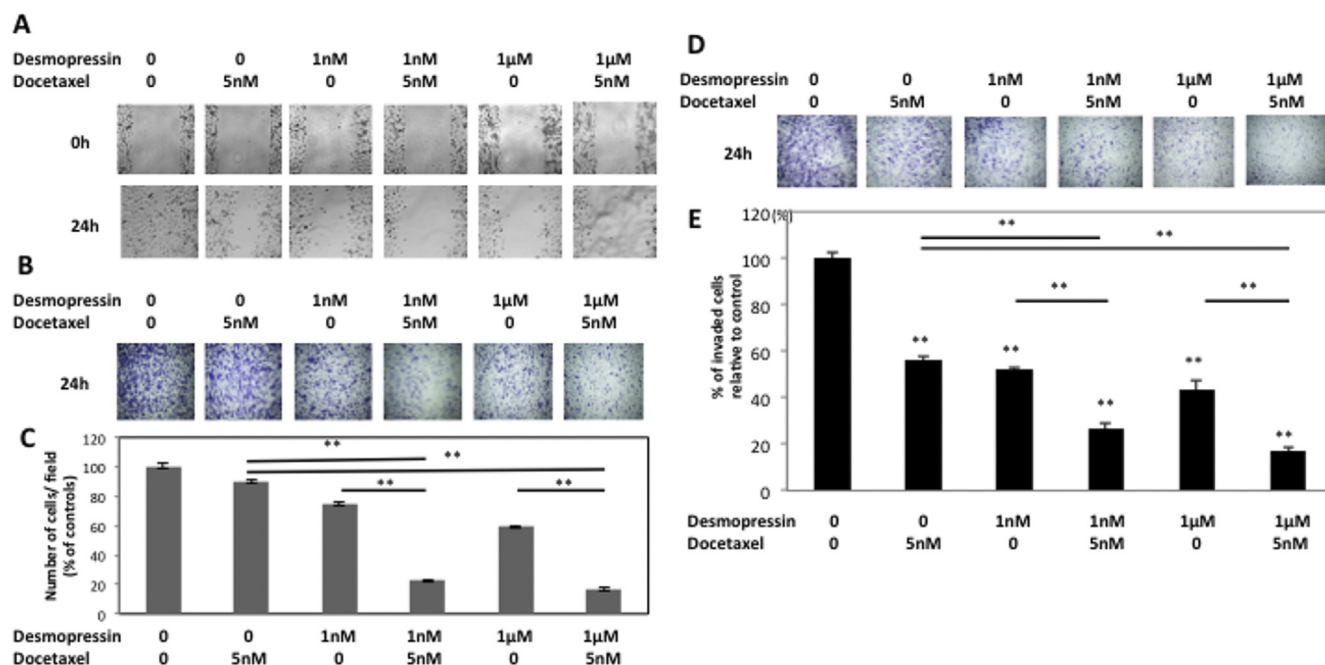
### 3.6. uPA-MMP pathway mediates the desmopressin-induced migration and invasion of prostate cancer cells

The two important molecules MMP and uPA are involved in cancer cell invasion, motility, and tumor dormancy [15–17]. Based on the results of both the migration assay and invasion assay in PC3 cells, we anticipated that desmopressin used as a monotherapy may have anti-metastatic potential. Dose standardization study was carried out with desmopressin. Desmopressin monotherapy altered the expression of precursor pro-uPA, active uPA, MMP-2, and MMP-9, all of which were reduced in a dose dependent manner (Supplementary Fig. 2). Desmopressin monotherapy reduced zymogen type uPA expression, thus attenuating uPA activity on the cell surface. These results demonstrate that desmopressin has the ability to inhibit tumor cell migration and invasion via the uPA-MMP pathway. We have looked at the expression (Western blotting) as well as the levels of activity of uPA, MMP-2 and MMP-9 (ELISA) following treatment with desmopressin and docetaxel. As shown in Fig. 3, combination treatment resulted in a more significant reduction in the expression of uPA when compared to the control (Fig. 3, A,  $^{**}p < 0.01$ ), and desmopressin monotherapy (Fig. 3, A,  $^{*}p < 0.05$ ). In addition, treatment with 5 nM docetaxel in combination with 1  $\mu$ M desmopressin showed a significant decrease in uPAR expression (Fig. 3, B  $^{*}p < 0.05$ ). The expression of MMP-2 was also significantly reduced in the cells treated with each combination therapy when compared to the control (Fig. 3, C,  $^{**}p < 0.01$ ), but not significantly different to each monotherapy. The expression of MMP-9 was significantly reduced in the cells treated with each combination therapy when compared to control and either of the monotherapy (Fig. 3, D,  $^{**}p < 0.01$ ). We found that decompression alone significantly inhibited the activity of uPA, MMP-2, and MMP-9 as determined by ELISA. Also, combination therapy resulted in decreased uPA, MMP-2, and MMP-9 expressions (Fig. 3, E, F and G). Consequently, these results clearly suggest that desmopressin in combination with docetaxel influences the uPA-MMP activity.

### 3.7. Desmopressin in combination with docetaxel inhibits PC-3 cell growth in a xenograft model

Tumors in control animals grew rapidly, measuring a volume of 923 mm<sup>3</sup> on day 35 post tumor inoculation. In contrast, tumor growth in the desmopressin-treated mice had a significantly slower rate of tumor development reaching a mean volume of 642 mm<sup>3</sup> on day 35, resulting in a 30.4% reduction when compared to the control (Fig. 4, Student's t-test  $p < 0.01$ ). The tumor volumes between the control and desmopressin treatment were significantly different (ANOVA,  $p < 0.01$ ). We went on to further investigate whether desmopressin was able to enhance the sensitivity of PC3 cells to docetaxel *in vivo*. As shown in Fig. 4, the tumors in the combination group were significantly smaller than either the control group, desmopressin treated group (ANOVA,  $p < 0.01$ ), or the docetaxel treated group (ANOVA,  $p < 0.05$ ). Statistical analysis confirmed that the combination treatment resulted in a significant inhibitory effect on tumor growth in the PC3 xenografts. Treatment with intravenous injection of docetaxel and/or desmopressin was well tolerated. All mice consistently maintained their body weight during each study.





**Fig. 2.** Desmopressin in combination with docetaxel treatment inhibits the migration and the invasion of PC3. (A) Wound healing assay, after 24 h, the numbers of cells were confluent in control group. Whereas, decreased cell numbers in docetaxel monotherapy, each desmopressin treatment groups and combination groups. (B) The migration of PC3 cells through a transwell filter, were assessed with or without 5 nM docetaxel in the presence of 1 nM or 1 μM of desmopressin. (C) Each combination therapy resulted in a significant reduction in cell migration compared to control and corresponding docetaxel alone as well as desmopressin alone group in the migration chamber assay (\*\*p < 0.01, Student's t-test). (D) The invasion of PC3 cells through a matrigel-coated transwell filter, were assessed with or without 5 nM docetaxel in the presence of 1 nM or 1 μM of desmopressin. (E) Invasion assay showed that each combination treatment of desmopressin and docetaxel resulted in a significant reduction in cell invasion when compared to the control and corresponding docetaxel alone as well as desmopressin alone group (\*\*p < 0.01, Student's t-test).

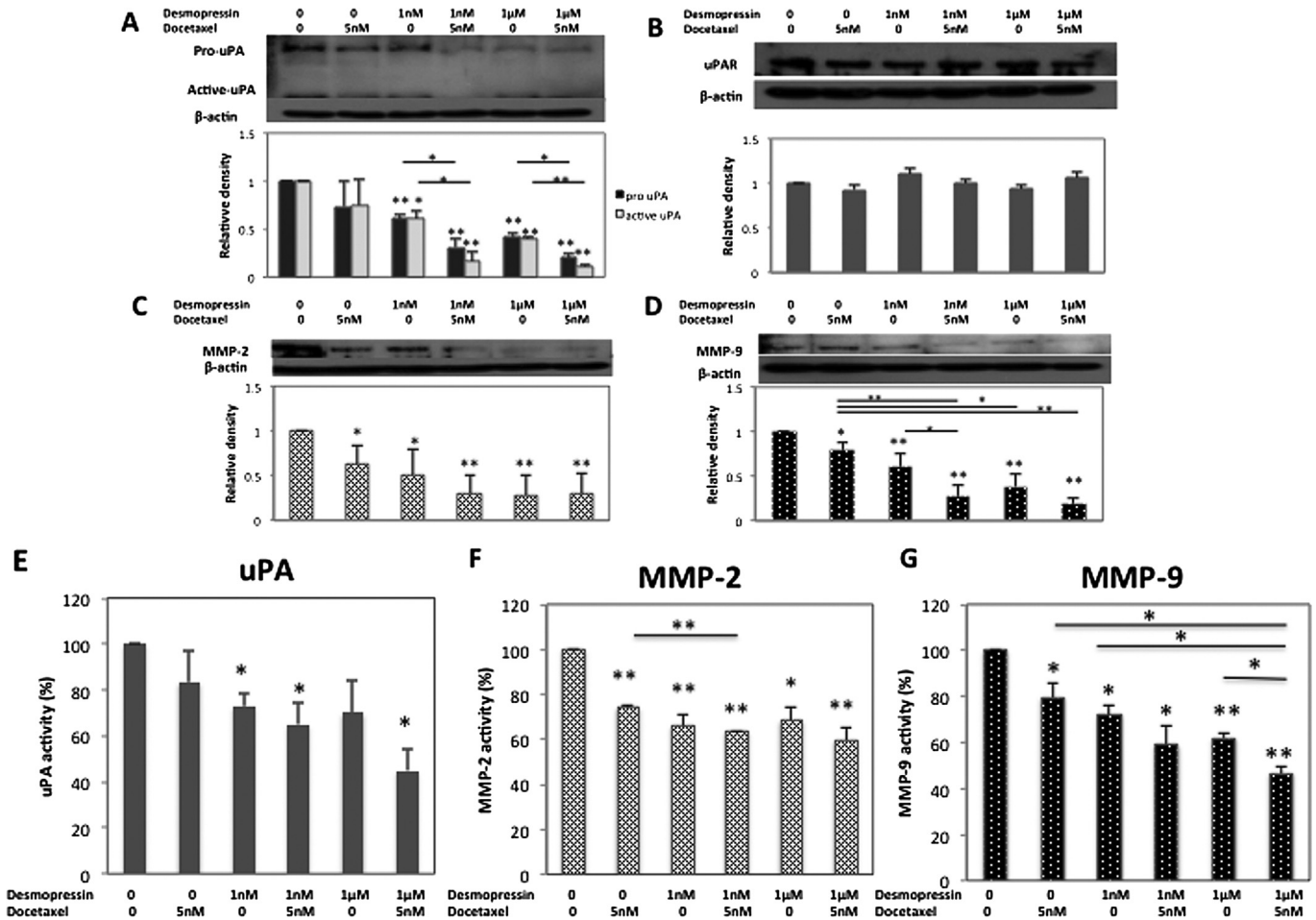
#### 4. Discussion

In this study we found desmopressin to have anti-proliferative, anti-migration, and anti-invasive effects on prostate cancer cells *in vitro* and *in vivo*. We are the first to report the mechanism of action of desmopressin in combination with docetaxel in a prostate cancer model. Our results have shown that in PC3 cells, both a low dose desmopressin (1 nM) as well as a higher dose (1 μM), have a significant anti-proliferative effect in *in vitro*. Desmopressin has different pharmacologic effects including anti-hemostatic and anti-diuretic effects, with higher doses having an anti-hemostatic effect [18]. Our results suggest that a high dose of desmopressin inhibited cell proliferation via the uPA-MMP pathway. The growth of prostatic tumor cells related to the activation of the plasminogen activator system is derived from the demonstration that growth rates and uPA production in tumor cells cultured at a low density are higher than those observed in cells grown at a higher cell density. This modulation may result in altered tumor cell proliferation [19]. Alteration in cellular proliferation was not dose dependent in comparison to the expression of uPA activity; which was decreased with increasing doses of desmopressin. It is possible that many cytokines and growth factors such as TGFβ1, IGF-1, FGF, EGF and bombesin induce the expression of components of the uPA system [15,20–22]. These factors may also be associated with uPA expression and tumor cell proliferation. Further studies are needed to elucidate the mechanism that causes the anti-proliferative effect for low-dose desmopressin in PC3 cells. Desmopressin monotherapy did not alter cell cycle distribution and the expression levels of apoptosis related proteins in both cell lines. Nonetheless, in PC3 cells, we observed that desmopressin, in combination with docetaxel, enhanced cell cycle arrest and the apoptotic effect. Our study

demonstrated that desmopressin functions as a docetaxel chemosensitizer for androgen independent cells. Further analysis could be needed to elucidate the mechanism that leads to the anti-tumor effect of desmopressin for androgen dependent cells.

We have shown that the anti-metastatic activity of desmopressin is mediated through the uPA pathway. The uPA and its receptor uPAR are expressed in most solid and invasive cancers including PC3 cells. However, LNCaP cell does not express the uPA and uPAR [19]. The uPA protein is involved in the degradation of the extracellular matrix, facilitating invasiveness and growth [23]. Our present study demonstrated desmopressin as monotherapy and in combination with docetaxel significantly inhibited the expression of uPA, MMP-2, and MMP-9. uPA activates MMP-2 and MMP-9 during the migration and invasion of prostate cancer [24]. The concept of the key role of the binding of uPA to uPAR derives from several studies demonstrating that the ability of tumor cells to invade and metastasize is downregulated by uPA inhibitors. Our results suggest that desmopressin monotherapy reduced pro-uPA expression as well as active uPA, thus attenuating uPA activity on the cell surface. Accordingly, desmopressin may be acting to inhibit uPA activity in prostate cancer.

One of the limitations associated with this study was the animal model. We have not performed *vivo* experiments to explain the anti-metastatic effects. Future studies could be conducted in transgenic animals to elucidate the potential anti-metastatic effects in an *in vivo* setting. In conclusion, this is the first study revealing the anti-tumor effect of desmopressin in combination with docetaxel for prostate cancer cells working activity. Our finding could potentially contribute to the therapeutic profile of desmopressin and enhance the efficacy of docetaxel based treatment for castrate resistant prostate cancer. Clinical studies of this innocuous combination are warranted.



**Fig. 3.** Western blot analysis was used to demonstrate the anti-metastatic effect of desmopressin used in combination with docetaxel study on PC3 cells. Blots showing levels of expression of (A) uPA, (B) uPAR, (C) MMP-2, and (D) MMP-9. Both combination therapies decreased uPA, MMP-2 and MMP-9 expressions (\* $p < 0.05$  and \*\* $p < 0.01$ , Student's t-test). Also activity assays were used to demonstrate the anti-metastatic effect of desmopressin used in combination with docetaxel study on PC3 cells. Assays showing levels of activity of (E) uPA, (F) MMP-2, and (G) MMP-9. Both combination therapies decreased uPA, MMP-2 and MMP-9 activities. (\* $p < 0.05$  and \*\* $p < 0.01$ , Student's t-test).

### Conflict of interest

The authors declare no conflict of interest.

### Acknowledgment

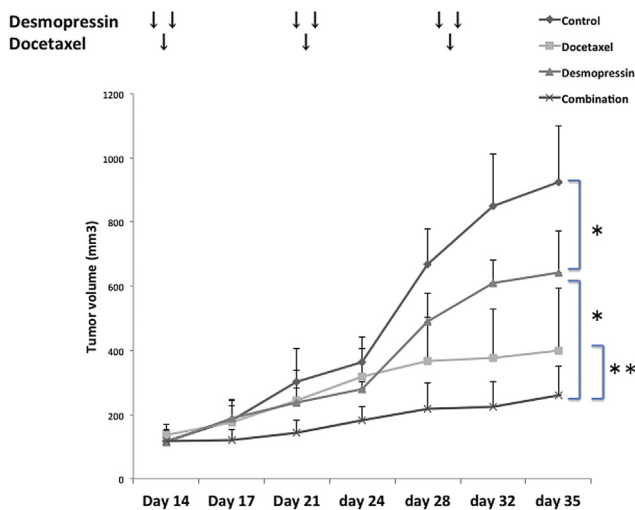
This work was supported by funds provided by Ferring Inc. (Canada). The funders had no role in this study design, data collection, analysis, and decision to publish. We would also like to thank the staff in the comparative research department, at the Sunnybrook Research Institute, for their technical support.

### Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.bbrc.2015.07.050>.

### Transparency document

Transparency document related to this article can be found online at <http://dx.doi.org/10.1016/j.bbrc.2015.07.050>.



**Fig. 4.** Effects of desmopressin and/or docetaxel treatments on growth of PC3 xenograft model. Nude mice were subcutaneously inoculated with  $1 \times 10^6$  PC3 cells per mouse. The tumor volumes were measured twice per week for 35 days (\* $p < 0.05$  and \*\* $p < 0.01$ , Student's t-test or ANOVA).

## References

- [1] D.P. Petrylak, C.M. Tangen, M.H. Hussain, et al., Docetaxel and estramustine compared with mitoxantrone and prednisone for advanced refractory prostate cancer, *N. Engl. J. Med.* 351 (2004) 1513–1520.
- [2] I.F. Tannock, R. de Wit, W.R. Berry, et al., Docetaxel plus prednisone or mitoxantrone plus prednisone for advanced prostate cancer, *N. Engl. J. Med.* 351 (2004) 1502–1512.
- [3] D. Friedland, J. Cohen, R. Miller Jr., et al., A phase II trial of docetaxel (Taxotere) in hormone-refractory prostate cancer: correlation of antitumor effect to phosphorylation of Bcl-2, *Semin. Oncol.* 26 (1999) 19–23.
- [4] D.W. Richardson, A.G. Robinson, Desmopressin, *Ann. Intern. Med.* 103 (1985) 228–239.
- [5] P.M. Mannucci, M. Aberg, I.M. Nilsson, et al., Mechanism of plasminogen activator and factor VIII increase after vasoactive drugs, *Br. J. Haematol.* 30 (1975) 81–93.
- [6] D.F. Alonso, G. Skilton, E.F. Farías, et al., Antimetastatic effect of desmopressin in a mouse mammary tumor model, *Breast Cancer Res. Treat.* 57 (1999) 271–275.
- [7] S. Giron, A.M. Tejera, G.V. Ripoll, et al., Desmopressin inhibits lung and lymph node metastasis in a mouse mammary carcinoma model of surgical manipulation, *J. Surg. Oncol.* 81 (2002) 38–44.
- [8] G.V. Ripoll, S. Giron, M.J. Krzymuski, et al., Antitumor effects of desmopressin in combination with chemotherapeutic agents in a mouse model of breast cancer, *Anticancer Res.* 28 (2008) 2607–2611.
- [9] V. Venkateswaran, L.H. Klotz, N.E. Fleshner, Selenium modulation of cell proliferation and cell cycle biomarkers in human prostate carcinoma cell lines selenium modulation of cell proliferation and cell cycle biomarkers in human prostate carcinoma cell lines, *Cancer Res.* 62 (2002) 2540–2545.
- [10] N.A. Venier, A.J. Colquhoun, H. Sasaki, et al., Capsaicin: a novel radio-sensitizing agent for prostate cancer, *Prostate* 75 (2015) 113–125.
- [11] A.J. Colquhoun, N.A. Venier, A.D. Vandersluis, et al., Metformin enhances the antiproliferative and apoptotic effect of bicalutamide in prostate cancer, *Prostate Cancer Prostatic Dis.* 15 (2012) 346–352.
- [12] C.-C. Liang, A.Y. Park, J.-L. Guan, In vitro scratch assay: a convenient and inexpensive method for analysis of cell migration in vitro, *Nat. Protoc.* 2 (2007) 329–333.
- [13] M.L. Immordino, P. Brusa, S. Arpicco, et al., Preparation, characterization, cytotoxicity and pharmacokinetics of liposomes containing docetaxel, *J. Control Release* 91 (2003) 417–429.
- [14] K.J. Pienta, Preclinical mechanisms of action of docetaxel and docetaxel combinations in prostate cancer, *Semin. Oncol.* 28 (2001) 3–7.
- [15] K. Forbes, K. Gillette, L.A. Kelley, et al., Increased levels of urokinase plasminogen activator receptor in prostate cancer cells derived from repeated metastasis, *World J. Urol.* 22 (2004) 67–71.
- [16] A. Moroz, F.K. Delella, L.M. Lacorte, et al., Fibronectin induces MMP2 expression in human prostate cancer cells, *Biochem. Biophys. Res. Commun.* 430 (2013) 1319–1321.
- [17] A.D. Kacsinta, C.S. Rubenstein, I.C. Sroka, et al., Intracellular modifiers of integrin alpha 6p production in aggressive prostate and breast cancer cell lines, *Biochem. Biophys. Res. Commun.* 454 (2014) 335–340.
- [18] J.E. Kaufmann, U.M. Vischer, Cellular mechanisms of the hemostatic effects of desmopressin (DDAVP), *J. Thromb. Haemost.* 1 (2003) 682–689.
- [19] C. Festuccia, V. Dolo, F. Guerra, et al., Plasminogen activator system modulates invasive capacity and proliferation in prostatic tumor cells, *Clin. Exp. Metastasis* 16 (1998) 513–528.
- [20] M. Bologna, C. Festuccia, P. Muzi, et al., Bombesin stimulates growth of human prostatic cancer cells in vitro, *Cancer* 63 (1989) 1714–1720.
- [21] D.F. Jarrard, B.F. Blitz, R.C. Smith, et al., Effect of epidermal growth factor on prostate cancer cell line PC3 growth and invasion, *Prostate* 24 (1994) 46–53.
- [22] K. Danø, P.A. Andreasen, J. Grøndahl-Hansen, P. Kristensen, et al., Plasminogen activators, tissue degradation, and cancer, *Adv. Cancer Res.* 44 (1985) 139–266.
- [23] P.H. Quax, A.C. de Bart, J.A. Schalken, et al., Plasminogen activator and matrix metalloproteinase production and extracellular matrix degradation by rat prostate cancer cells in vitro: correlation with metastatic behavior in vivo, *Prostate* 32 (1997) 196–204.
- [24] C. Festuccia, D. Giunciuglio, F. Guerra, et al., Osteoblasts modulate secretion of urokinase-type plasminogen activator (uPA) and matrix metalloproteinase-9 (MMP-9) in human prostate cancer cells promoting migration and matrigel invasion, *Oncol. Res.* 11 (1999) 17–31.